Vaccine-delivered HIV envelope inhibits CD4+ T-cell activation, a mechanism for poor HIV vaccine responses

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The human immunodeficiency virus (HIV) causes impairment of the immune system in part by targeting CD4+ T cells for infection and dysfunction. HIV envelope (Env) protein present on free virions and infected cells causes dysfunction of uninfected bystander CD4+ T cells via interaction with both CD4 and coreceptors. Env is commonly used as part of a cocktail of HIV antigens in current vaccines. In DNA and viral vector vaccine approaches, antigen-presenting cells (APCs) and non-APCs in the vicinity of the vaccine delivery site and draining lymph node express vaccine-derived antigens. The studies here demonstrate that cell-surface expression of Env on APCs and non-APCs as part of the vaccine action causes an inhibition of antigen-induced CD4+ T-cell activation and proliferation mediated by CD4 binding and suggests a potential mechanism for reduced activity of Env-containing HIV vaccines. Similar studies using a functional Env lacking CD4 binding circumvented suppression, suggesting an alternative and potentially superior approach to HIV vaccine design. (Blood. 2007;109:2538-2544)

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Introduction

It is estimated that 4.9 million new HIV infections occurred in 2005, mostly in regions of the world with limited access to health care. The development of a vaccine is the most effective approach to combat this epidemic. The envelope (Env) protein of the virus is a key component of an effective vaccine because it exclusively induces neutralizing antibodies, which are protective in the macaque model. However, Env has been demonstrated to suppress CD4+ T cells by multiple mechanisms. Inhibition has been characterized by a lack of proliferation, the induction of energy, and various apoptotic mechanisms. The role of Env interactions with CD4 and coreceptors in mediating the various forms of suppression is under debate with studies demonstrating a role for each. R5 Env caused apoptosis of CD4+ T cells from a CCR5Δ32 homozygous donor, whereas, in another study, Env interaction with CXCR4 and a nonsignaling CD4 induced CD4+ T-cell apoptosis. The mechanism of suppression observed and the receptors involved varied according to the nature of Env used and the cell culture system studied. Env is a trimeric protein with each of its monomers possessing a CD4 binding site. Thus, an Env trimer or aggregates created by antibodies against monomeric gp120 subunits can mediate cross-linking of CD4 and/or coreceptor leading to signaling. In patients infected with HIV, trimeric Env capable of suppression is found on the surface of infected cells as well as on the virus itself. The use of Env in HIV vaccines, especially in commonly used DNA and viral vectors, could be envisioned to deliver similar CD4+ T-cell suppressive signals.

Although antigen-presenting cells (APCs) such as dendritic cells (DCs) are prime targets for vaccines, other cell types at the site of vaccine delivery, and potentially the major vaccine antigen-presenting cells types, are non-APCs such as myocytes, fibroblasts, subepithelial, and epithelial cells for muscle, dermal, subcutaneous, and mucosal delivery, respectively, as well as cells in the draining lymph nodes. CD4+ T cells that are recruited to the site of vaccine delivery as well as to the draining lymph node in response to vaccine-induced inflammation may then interact with and be suppressed by interaction with Env expressed on the surface of all of these cells.

We designed an in vitro experimental model to mimic in vivo Env vaccine delivery. Our results demonstrate that APCs and non-APCs expressing Env of both R5 and X4 coreceptor usage mediated suppression of CD4+ T-cell activation and proliferation in response to antigen. In addition, we identify model X4 and R5 nonsuppressive Env, which mediate fusion and infection, thus preserving important neutralizing epitopes, and present a new approach for more effective Env immunogens in vaccine design.

Materials and methods

Cell culture

Peripheral blood mononuclear cells (PBMCs) from HIV-seronegative donors were obtained through an IRB-approved protocol. Approval was obtained from the University of Pennsylvania institutional review board for these studies. Informed consent was provided according to the Declaration of Helsinki. CD4+ T cells were purified by negative selection using the Human T-cell Subset Column kit (R & D Systems, Minneapolis, MN). Human embryonic kidney 293 cells (ATCC, Gaithersburg, MD) were propagated in DMEM supplemented with glutamine (Invitrogen, Carlsbad, CA) and 10% FCS (HyClone, Ogden, UT) (complete medium). DCs were produced as described previously and treated with GM-CSF (50 ng/mL) + IL-4 (100 ng/mL) (R & D Systems) in AIM V medium (Invitrogen). In certain experiments, PBMCs or purified CD4+ T cells were stained with 3.33 µM CFSE (Molecular Probes, Carlsburg, CA) according to the manufacturer’s protocol.
Plasmid constructs and RNA synthesis

Env genes from plasmids encoding IIIB, 8X, 8XD, 8XV3Bal, and 8XDv3Bal Env18–20 were cloned into the RNA expression plasmid T7TS.21 Plasmids PCR3-89.6, pcDNA3-SF162, PCI-YU-2, and pcDNA3-JR-F1 containing upstream T7 promoter sites were obtained from the AIDS Reference and Reagent Program (Bethesda, MD). Plasmids were linearized at unique sites downstream of the 3′ coding sequence and in vitro transcribed with T7 polymerase to synthesize m7GpppG-capped mRNA with additional poly(A) tail using the mMessage T7 ultra kit (Ambion, Austin, TX). RNA was complexed to lipofectin and delivered to 293T cells as described.22

Cocultures of Env-expressing 293T cells and CFSE-labeled cells

293T cells in 6- or 12-well plates were transfected with Env or luciferase encoding mRNA or poly(A, C) RNA (Sigma, St Louis, MO), incubated for 4 to 5 hours, and cocultured with 4 or 2 × 10⁶ CFSE-labeled PBMCs or negatively selected purified CD4⁺ T cells with 0.1 µg/ml superantigen TSST-1 (toxic shock syndrome toxin-1; Sigma) or anti-CD3 mAb (0.5-1 µg/ml; Muromonab-CD3; Ortho Biotech, Bridgewater, NJ). In certain experiments, 2 µg/ml monomeric IIIB or SF162 gp-120, 20 to 80 U/ml IL-2 (AIDS Reference and Reagent Program), and/or 10 µg/ml sCD4 (Progenics, Tarrytown, NY) were added. Proliferation of CD4⁺ T cells by CFSE dilution was assayed by flow cytometry 4 to 5 days after initiation of coculture. CD4⁺ T cells were identified using anti-CD4 antibody conjugated to allophycocyanin (BD Pharmingen, San Jose, CA) or mouse anti-human CD4 (Clone OKT4) and goat anti-mouse Ig-allophycocyanin as a secondary antibody.

Detection of apoptosis and activation in CD4⁺ T cells exposed to Env

PBMCs were cocultured with Env-expressing 293T cells or 700 µM cycloheximide (CHX; Sigma) and activated with 0.1 µg/ml TSST-1 or 1 µg/ml anti-CD3 mAb. Apoptosis was examined 6, 17, and 36 hours after coculture and identified as annexin V–positive (BD Pharmingen, San Diego, CA) and 7-AAD–negative (Sigma) cells. Total cell death was measured by positive staining for 7-AAD in ungated cells. For studies measuring up-regulation of activation markers in Env-treated PBMCs, CD69 peridinin chlorophyll protein (day 1), HLA-DR fluorescein isothiocyanate and CD25 phycoerythrin cyanine 7 (BD Immunocytometry Systems, San Jose, CA) (day 2) were used.

Purified CD4⁺ T cells were exposed to IIIB Env-expressing 293T cells for 1 day, stimulated with 1 µg/ml anti-CD3 antibody for 10 minutes for p38 or 2 days followed by 20 U/ml IL-2 for 2 to 20 minutes for STAT5a, fixed, and analyzed by flow cytometry using anti–phospho-MAPK (pT180/p38 or 2 days followed by 20 U/ml IL-2 for 2 to 20 minutes for STAT5a, fixed, and analyzed by flow cytometry using anti–phospho-AKT (ser 473) Alexa Fluor488 (Cell Signaling Technologies, Danvers, MA) and anti-CD4 v4 (clone L120) phycoerythrin (BD Immunocytometry Systems).

DC infection with vaccinia virus

DCs were suspended at a concentration of 2 million/mL and infected with vaccinia-encoding IIIB, MN, or 921-U5037.8 Env (NIH AIDS Research and Reference Reagent Program), T7 Polymerase (VTF7.3),23 or with wild-type WR strain for 2 hours, washed, and incubated for 4 hours. Infected DCs were cocultured with CFSE-labeled PBMCs at a ratio of 1:10 and activated with 0.1 ng/ml TSST-1.

Protein expression by fusion assay and Western blot

The protocol previously used by Nussbaum et al24 and Rucker et al25 was used for fusion assays between mRNA-encoding Env-transfected 293T cells and Q76 cells expressing CD4 and/or CXCR4 or CCR5. For Western blot analysis, 25 µg protein in lysates from Env- or luciferase-transfected 293T cells were analyzed using rabbit polyclonal anti-IIIB Env serum.26

Results

Env induces suppression of CD4⁺ T-cell proliferation

Following vaccine delivery, a number of cell types other than APCs take up and express vaccine-derived antigens. Expression by these cells presents Env in the absence of costimulatory molecules. 293T cells were transfected with Env-encoding mRNA as consistent, and low-level protein expression is obtained in most cells,17 allowing uniform exposure of CD4⁺ T cells to Env. PBMCs were used as a source of CD4⁺ T cells and cocultured with 293T cells expressing X4-tropic IIIB, dual tropic 89.6, and M-tropic JR-FI, YU-2, and SF162 EnvS. Cells were stimulated with a concentration of anti-CD3 mAb that typically stimulates one quarter of the CD4⁺ T cells. To monitor proliferation, PBMCs were stained with CFSE (5-and 6-carboxyfluorescein diacetate, succinimidyl ester),27 CD4⁺ T cells that encountered any of the EnvS had lower proliferation levels compared with cultures exposed to control RNA-transfected 293T cells (Figure 1A). Similar inhibition was observed when cells were activated by the superantigen TSST-1, a model antigen for activation via the T-cell receptor (TCR), which requires the same

Figure 1. HIV Env suppresses anti-CD3-activated CD4⁺ T-cell proliferation. (A) 293T cells expressing IIIB, 89.6, YU-2, SF162, and JR-FI EnvS or no Env (poly(A, C) RNA transfected) were cocultured with CFSE-labeled PBMCs with or without sCD4. CD4⁺ T cells were activated with anti-CD3 mAb (0.5 µg/mL), stained with anti-CD4 mAb, and monitored for proliferation 4 days after coculture by flow cytometry. (B) Negatively selected purified CD4⁺ T cells were cocultured with 293T cells expressing IIIB Env or no Env and activated using anti-CD3 mAb (0.5 µg/mL). Fusion assays demonstrated surface expression of functional Env. Encoding mRNA-transfected 293T cells were mixed with Q76 cells expressing CD4 and CXCR4 (C) or CCR5 (D), and fusion of the 2 cell populations was measured by luciferase production. Quadrants are labeled with the percent of events in each. Experiments shown are representative of 3 independent experiments.
costimulatory and adhesion molecules as conventional anti-
gens.25,29 This suggested that suppression of proliferation was not
dependent on a particular mode of activation. In addition, nega-
tively selected purified CD4+ T cells exposed to Env also failed to
proliferate, arguing against a role for CD8+ T cells and natural
killer cells in mediating suppression (Figure 1B).

To characterize the signal delivered by Env, we examined
whether suppression could be overcome by blocking Env interac-
tion with CD4. Soluble (s)CD4 almost completely abrogated
Env-induced suppression (Figure 1A). P values were calculated
using results from 3 independent experiments. A paired Student t
test with 2 tails was performed using the percentage of change from
total CD4+ T cells.

Expression of functional trimeric Env on 293T cells was
confirmed in fusion assays demonstrating that mRNA delivery
resulted in surface expression of fusion-competent Env (Figure 1C).
Delivery of Env-containing vaccines in vivo results in a
variety of expression levels that are governed by the cellular
vaccine target and the vaccine delivery method. In our experimen-
tal system that mimics vaccination with Env, we chose delivery
of Env by encoding mRNA to 293T cells, which results in expression
of Env in nearly 100% of cells at much lower levels compared with
other methods of delivery, such as DNA. This was important as
lymphocyte trafficking does not occur in our system. Thus, if only
25% of the cells expressed Env, then only about 25% of the CD4+ T
cells added would be exposed. This low level of expression was
sufficient to mediate suppression, and, in fact, a 5-fold reduction in
the quantity of Env encoding mRNA delivered which results in a
5-fold lower level of protein expressed resulted in similar levels of
suppression (data not shown). Thus, at the range of expression
levels seen in our culture system significantly large variations in the
amount of Env expressed did not alter the degree of suppression
everted. This could explain the lack of correlation observed
between the degree of suppression mediated by an Env strain
(Figure 1A) and the amount of fusion mediated (Figure 1C-D).

Several reports have found that trimeric or aggregated forms
of Env are required to mediate suppression.5,8 However, others have
found that monomeric gp120 could exert suppressive effects.3,7 In

-Table 2. Effect of monomeric gp120 on CD4+ T-cell proliferation

<table>
<thead>
<tr>
<th></th>
<th>No Env</th>
<th>IIIB gp120</th>
<th>SF162 gp120</th>
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<tr>
<td>CD4+ cells undergoing proliferation, %</td>
<td>15.3</td>
<td>14.4</td>
<td>12.5</td>
</tr>
<tr>
<td>CD4+ cells undergoing proliferation, %</td>
<td>7.7</td>
<td>7.4</td>
<td>7.3</td>
</tr>
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Anti-CD3 mAb (0.5 μg/mL) stimulated CFSE-labeled PBMCs were exposed to 2 μg/mL IIIB or SF162 gp120 or left untreated. CD4+ T cells were monitored for proliferation 4 days after coculture by flow cytometry and expressed as percentage of total CD4+ T cells.

-Table 3. Statistical analysis of suppression of CD4+ T cells

<table>
<thead>
<tr>
<th>Env strain</th>
<th>P of suppression compared with no Env control</th>
<th>P of suppression compared with sCD4 treatment</th>
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<tbody>
<tr>
<td>IIIB</td>
<td>&lt;.005</td>
<td>&lt;.005</td>
</tr>
<tr>
<td>89.6</td>
<td>.01</td>
<td>.02</td>
</tr>
<tr>
<td>JR-F1</td>
<td>.02</td>
<td>.04</td>
</tr>
<tr>
<td>Yu-2</td>
<td>.01</td>
<td>.02</td>
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P values were calculated using results from 3 independent experiments. Percentage of change from control (no Env) or sCD4 treatment was used in a paired Student t test with 2 tails.

Figure 2. DCs expressing Env and presenting antigen suppress CD4+ T-cell proliferation. DCs were infected with vaccinia virus encoding IIIB Env or control vaccinia encoding T7 polymerase and cocultured with CFSE-labeled PBMCs with 0.1 ng/mL TSST-1 and with or without sCD4. This concentration of superantigen requires presentation by DCs as no proliferation is observed in their absence. Proliferation of CD4+ T cells was measured 5 days later by flow cytometry. Quadrants are labeled with the percent of events in each. Experiments shown are representative of 3 independent experiments.
proliferation. In addition, the ability of Env to interact with CD4 molecules on DCs and suppress their maturation and function cannot be ruled out in our system as it has been observed in other systems. Expression of functional Env was confirmed in fusion assays between DCs and Quail cells (data not shown). Thus, mature DCs expressing Env mediate suppression of antigen-specific CD4+ T-cell proliferation despite expressing multiple costimulatory molecules.

Neither apoptosis nor IL-2–dependent anergy mediate CD4+ T-cell suppression

Env-induced CD4+ T-cell dysfunction has been described as anergy, apoptosis, cell death, or a failure to proliferate. We observed that Env suppresses CD4+ T cells in our system by hindering proliferation. We next investigated whether anergy or apoptosis occurred. PBMCs were cocultured with Env-expressing 293T cells and were activated using TSST-1 superantigen. Six, 18, and 36 hours later, CD4+ T cells were analyzed for early apoptosis using annexin V staining of 7-AAD–negative cells. CHX is a protein synthesis inhibitor that induces apoptosis of PBMCs and was used as a positive control. Env failed to induce apoptosis of CD4+ T cells above the level seen in cells unexposed to Env (Figure 3A). In addition, an increase in cell death (7-AAD uptake in un gated cells) in Env-exposed CD4+ T cells was not observed (Figure 3B). Thus, apoptosis and cell death are not major contributors to the suppression of proliferation induced by Env in our system.

T-cell anergy is defined as a state of nonresponsiveness to activation with antigen. The addition of exogenous IL-2 overcomes most but not all types of anergy. Selliah and Finkel observed that CD4+ T-cell suppression induced by Env could be overcome with exogenous IL-2. To investigate this mechanism, CFSE-stained PBMCs were cultured with control or Env-expressing 293T cells and activated with TSST-1 in the presence of increasing amounts of exogenous IL-2 (20, 40, or 80 U/mL). IL-2 was unable to prevent suppression of proliferation of CD4+ T cells by Env (Figure 3C; data not shown), arguing against this form of anergy as the mechanism of suppression.

CD4+ T cells activated in the presence of Env phosphorylate AKT, MAPK, and STAT5a but fail to completely up-regulate activation markers

CD4+ T cells exposed to Env failed to proliferate in response to antigen but did not apoptose, anergize, or die. We next sought to determine the location of the block in the activation cascade. In response to T-cell activation, phosphorylation of key signaling molecules p38 MAPK and AKT relay the TCR signal to downstream mediators. CD4+ T cells exposed to Env and activated by anti–CD3-phosphorylated AKT and MAPK similar to control-treated cells (Figure 4). IL-2 is an important cytokine involved in T-cell proliferation, and its binding to both the low- and high-affinity IL-2Rs leads to the phosphorylation of STAT5a. Exogenous IL-2 was unable to circumvent the suppression of proliferation induced by Env (Figure 3C), but cells exposed to and suppressed by Env were able to phosphorylate STAT5a in response to IL-2 (Figure 4), suggesting that early signaling mediators in the IL-2 cascade also were not impaired.

On activation, CD4+ T cells go through a series of events prior to proliferation, including up-regulation of markers of activation such as CD69, HLA-DR, and CD25. PBMCs were exposed to IIIB En-expressing 293T cells and activated using anti-CD3 or superantigen. Up-regulation of CD69 (day 1) and HLA-DR and CD25 (day 2) was monitored using flow cytometry. CD4+ T cells exposed to Env were unable to completely up-regulate their activation markers (Table 3; data not shown). Thus, although exposure of CD4+ T cells to Env did not suppress the phosphorylation of immediate signaling mediators, it suppressed the up-regulation of early and late cell-surface markers of activation and proliferation.

Functional Envs lacking CD4 binding do not suppress antigen-induced CD4+ T cells

Env-induced suppression could be overcome by preventing its interaction with CD4 (Figure 1A). We hypothesized that an Env lacking the ability to bind CD4 could serve as a better immunogen by circumventing CD4+ T-cell suppression. The chemokine receptor binding site is an important neutralization region. Thus, mutations that abrogate coreceptor binding, such as a loss of
Table 3. Effect of HIV Env on activation marker expression

<table>
<thead>
<tr>
<th>CD4+ cells expressing, %</th>
<th>No Env</th>
<th>IIIB Env</th>
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<tr>
<td>CD69</td>
<td>78.9</td>
<td>56.6</td>
</tr>
<tr>
<td>HLA-DR</td>
<td>23.5</td>
<td>12.9</td>
</tr>
<tr>
<td>CD25 (IL2R-α)</td>
<td>60.4</td>
<td>31.6</td>
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</tbody>
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<table>
<thead>
<tr>
<th>CD4+ cells expressing, %</th>
<th>No Env</th>
<th>IIIB Env</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD69</td>
<td>59.2</td>
<td>61.8</td>
</tr>
<tr>
<td>HLA-DR</td>
<td>43.7</td>
<td>41.6</td>
</tr>
<tr>
<td>CD25 (IL2R-α)</td>
<td>33.3</td>
<td>22.2</td>
</tr>
</tbody>
</table>

PBMCs were exposed to control or IIIB Env expressing 293T cells and activated with anti-CD3. Expression of CD69 on day 1 and HLA-DR and CD25 on day 2 on CD4+ and CD4+ T cells was determined using flow cytometry.

Discussion

The quest for an effective HIV vaccine faces many hurdles, including difficulty in developing strong and broad immune responses and the ability of the virus to evade immune recognition. The system studied here models the encounter of CD4+ T cells with both APCs and non-APCs expressing Env delivered as a vaccine. Both X4 and R5 Envs suppressed the activation and proliferation of CD4+ T cells with specific interaction with CD4. Suppression was characterized by a lack of proliferation despite normal activation of the main signaling pathways involved in T-cell activation and proliferation, AKT, MAPK, and STAT5a but a deficient up-regulation of early and late cell-surface activation markers. The suppression of antigen-induced proliferation was not due to IL-2-dependent anergy, apoptosis, or cell death.

The role of CD4 versus the coreceptors CXCR4 and CCR5 in mediating Env-induced suppression is controversial. Some reports have suggested a role for coreceptors in mediating suppression in conjunction with CD4. Others have suggested that Env can mediate suppression solely via CD4 or coreceptors. In this study, we show that Env delivered as a vaccine mediates a suppression of activation and proliferation regardless of its coreceptor usage. Although CD4 interaction is required for this suppression, both coreceptors could have played a role. To test this, we examined suppression of X4 Envs in the presence of AMD3100, which blocks Env interaction with CXCR4. AMD3100 caused a nonspecific increase in proliferation in both Env- and control-transfected 293T-exposed CD4+ T cells, which may be attributed to blocking inhibitory chemokine signaling through CXCR4. Similar studies with TAK-779, which blocks R5 Env interaction with CCR5, did not overcome suppression. This suggests that in our system studying vaccine-delivered Env in the absence of HIV infection, inhibition of CD4+ T-cell proliferation occurred in a CD4-dependent, chemokine receptor-independent manner.

Previous studies demonstrating the suppressive effects of Env have focused on a single aspect of CD4+ T-cell suppression such as apoptosis or anergy and used systems to model interactions that occur during HIV infection. In this report, we present a comprehensive study examining multiple facets of Env-induced suppression when it is used as an immunogen. DCs that acquire the vaccine will express protein from all the delivered antigens. When the protein is processed and presented to antigen-specific CD4+ T cells by the

Figure 5. Functional Envs lacking CD4 binding do not suppress antigen-induced CD4+ T-cell proliferation. 293T cells were transfected with 8X or 8XD Env-encoding mRNA; (A) 8XV3Bal or (B) 8XDV3Bal Env-encoding mRNA, and control poly(A) RNA and cocultured with CFSE-labeled PBMCs. CD4+ T cells were activated with 0.1 μg/mL TSST-1, and proliferation was measured 4 days later using flow cytometry. P values were .01 (A) and less than .005 (B) for comparison to sCD4 treatment. P values were less than .005 in comparing 8XD and 8XDV3Bal with no Env control. Quadrants are labeled with the percent of events in each. (C) Expression of Env protein by mRNA-transfected 293T cells was confirmed by Western blot analysis. (D) Surface expression of functional Env on mRNA-transfected 293T cells was determined using fusion assays. Error bars represent the standard error of the mean. Experiments shown are representative of 3 independent experiments.
APCs, suppression by surface-expressed Env would occur, thus hindering immune responses to all antigens in the cocktail. This is the first study demonstrating that Env when used as a vaccine could suppress immune responses, making the findings highly relevant to the field of vaccine design. Because Env is an essential component in an HIV vaccine that aims to generate neutralizing antibodies, inclusion of nonsuppressive forms of Env will enhance immune responses.

DCs are prime targets for vaccines in vivo. Studies have shown a good correlation between the uptake of vaccine by DCs and the magnitude of the immune response induced. The studies demonstrated here suggest that DCs that acquire an Env-containing vaccine could actually hinder the immune response. This hypothesis is supported by a nonhuman primate study where animals were immunized with identical Gag and Pol antigens in the presence or absence of Env using a vaccine regimen of plasmid DNA prime and replication-incompetent adenoviral vector boost. Inclusion of Env in the cocktail led to a decrease in Gag- and Pol-specific IFN-γ responses in PBMCs.

We describe a modified Env that circumvents this suppression without losing important neutralizing epitopes. The X4 Env 8XD and the R5 8XDV3Bal have 2 key advantages as vaccine models. First, they have a single amino acid loss of function mutation in their CD4 binding site, thereby preventing suppression. Second, they have a comparatively open conformation with fewer glycosylation sites, thereby stably exposing their chemokine receptor binding site. Exposure of this important neutralization site could induce a better directed neutralizing antibody response against this conserved site. These specific Envs would not become a component of a new vaccine approach, because effective vaccines would include primary isolates Env of suitable Clade and coreceptor specificity. The production of such an Env that shares these properties can be easily accomplished. HIV strains possessing the appropriate Env can be serially passaged in cell lines expressing decreasing amounts of CD4 to generate a CD4-independent variant. Env can then be cloned from the CD4-independent strain and mutated at a single amino acid (368 D to R) to abrogate CD4 binding. This results in an Env of suitable Clade and coreceptor specificity which is nonsuppressive because it lacks CD4 binding and should also stably expose its coreceptor binding site by virtue of its CD4 independence, possibly leading to enhanced neutralizing antibody responses.

HIV is an immunosuppressive virus and mediates its immune destructive activities in a number of manners. This includes direct killing of T helper cells, dysfunction of APCs and bystander T cells, and direct and indirect effects on almost all arms of the innate and acquired immune system. One major difficulty in developing an effective HIV vaccine is the lack of immune correlates of protection. A second that we demonstrate in this study is that immunogens, including Env used in HIV vaccines, are intrinsically immunosuppressive. We demonstrate that HIV Env, the only antigen capable of inducing neutralizing antibodies, inhibits the proliferation of CD4+ T cells in response to antigen. This inhibition to T helper cell proliferation is independent of the antigen stimulating the CD4+ T cells but occurs when the APCs or other cells in the region of the delivered vaccine take up the vaccine and express Env. Using an Env that is mutated to prevent CD4 binding but stably expresses the coreceptor binding site avoided this T-cell dysfunction. Model animal studies to determine whether this vaccine approach results in more potent immune responses are needed.

Acknowledgments

HIV-1 IIIB gp120; HIV envelope plasmids PC3-89.6, pcDNA3-SF162, PCI-YU-2, and pcDNA3-JR-F; and vaccinia viruses vP1198, vP1489, and vP1174 (Virogenic Corp) were obtained through the NIH AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, NIH. The polyclonal anti-IIIB Env rabbit serum and the vaccinia vTF7-3 were kind gifts from Dr Robert Doms and Dr Stuart Isaacs, University of Pennsylvania.

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Authorship

Contribution: K.F. designed and performed the research, analyzed the data, and wrote the paper; H.H. performed the research and analyzed the data; H.N. performed the research; J.A.H. designed the research and contributed vital new reagents; D.W. designed the research, analyzed the data, and wrote the paper.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

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References


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