Transforming growth factor-β1 modulates responses of CD34+ cord blood cells to stromal cell-derived factor-1/CXCL12

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Disruption of stromal cell-derived factor-1 (SDF-1/CXCL12 [CXC chemokine ligand 12]) interaction leads to mobilization of stem/progenitor cells from bone marrow to circulation. However, prolonged exposure of CD34+ cells to SDF-1 desensitizes them to SDF-1. So how do cells remain responsive to SDF-1 in vivo when they are continuously exposed to SDF-1? We hypothesized that one or more mechanisms mediated by cytokines exist that could modulate SDF-1 responsiveness of CD34+ cells and the desensitization process. We considered transforming growth factor-β1 (TGF-β1) a possible candidate, since TGF-β1 has effects on CD34+ cells and is produced by stromal cells, which provide niches for maintenance and proliferation of stem/progenitor cells. TGF-β1 significantly restored SDF-1-induced chemotaxis and sustained adhesion responses in cord blood CD34+ cells preexposed to SDF-1. Effects of TGF-β1 were dependent on the dose and duration of TGF-β1 pretreatment. Phosphorylation of extracellular signal-regulated kinase 1 (Erk1)/Erk2 was implicated in TGF-β1 modulation of migratory and adhesion responses to SDF-1. Our results indicate that low levels of TGF-β1 can modulate SDF-1 responsiveness of CD34+ cells and thus may facilitate SDF-1-mediated retention and nurturing of stem/progenitor cells in bone marrow. (Blood. 2005;106:485-493)

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Introduction

Hematopoiesis is a complex process whereby proliferation of hematopoietic stem/progenitor cells (HSPCs) ensures continuous renewal of mature blood cells. Stem cells engraft bone marrow following lethal irradiation/chemotherapy and reconstitute hematopoietic and immune compartments. Homing of HSPCs to marrow requires concerted action among factors essential for cellular migration, such as adhesion molecules and their ligands as well as chemotactic factors that lead them to a microenvironment for their growth and survival. In the bone marrow it is important that cells lodge in proper niches. Their retention depends on adhesive interactions between HSPCs and marrow stroma. The CXC chemokine receptor 4 (CXCR4)–stromal cell-derived factor-1 (SDF-1)/CXC chemokine ligand 12 (CXCL12) axis plays a critical role in homing and retention of HSPCs in the bone marrow.

SDF-1/CXCL12 is a potent chemotacticant for primitive and more mature hematopoietic cells. Stromal cells are potent sources of SDF-1. SDF-1 plays a key role during ontogeny of hematopoietic system. In adults, it promotes engraftment of transplanted HSPCs and hematopoietic reconstitution. The chemotactic effect of SDF-1 is mediated through the G protein-coupled receptor CXCR4. CXCR4, upon ligand binding, activates integrin-mediated firm adhesion and transmigration of HSPCs through bone marrow endothelium and stromal cells.

Rapid desensitization is a hallmark of G-protein–coupled receptors, leading to transient responses of short duration. During steady-state hematopoiesis, CXCR4/SDF-1 interactions restrict HSPCs in marrow and disruption of the SDF-1/CXCR4 axis leads to their egress into the circulation. This raises an important question. How do HSPCs remain responsive to SDF-1 in vivo when HSPCs are being continuously exposed to SDF-1? We considered the possibility that responses of HSPCs to SDF-1 could be modulated by other cytokines, and one of these may be transforming growth factor-β1 (TGF-β1).

TGF-β1, a pleiotropic factor, plays an important role in regulating the balance between proliferation and differentiation of hematopoietic cells. TGF-β type I and type II receptors are expressed on primitive progenitors and throughout stages of their maturation and development. Like SDF-1, TGF-β1 is produced by stromal cells. TGF-β1 induces chemotaxis of human mast cells, monocytes, and monocyctic dendritic cells and increases expression of integrin receptors very late antigen-4 (VLA-4) and VLA-5 on many cell types.

We examined the effect of prolonged exposure of CD34+ cells to SDF-1 on their chemotactic and adhesion responses to further SDF-1 stimulation and whether these responses could be modulated by TGF-β1. Our results demonstrate cross talk between TGF-β1 and SDF-1–CXCR4 interactions and suggests one means by which CD34+, CD34+CD38−, and myeloid progenitor cells may retain responsiveness to SDF-1 even in the presence of sustained exposure to SDF-1.

Materials and methods

Antibodies and reagents

The following antibodies were used for cell staining: fluorescein isothiocyanate (FITC)–conjugated CD34 (Miltenyi Biotec, Auburn, CA), phycoerythrin (PE)–conjugated CXCR4, and allophycocyanin (APC)–conjugated VLA-5. HSPCs were stained with PE–conjugated CD34 and APC–conjugated CD90 (Th17, CD90−/CD34+) evaluated as a proportion of the CD34+ population. A negative control, PE and APC–conjugated CD90−/CD34+ was used to determine background staining.

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CD34 (BD Pharmingen, San Diego, CA). For blocking integrin-mediated adhesion we used anti–VLA-4 (Clone HP1/2; Beckman Coulter, Fullerton, CA) and anti–VLA-5 (clone II A1; BD Pharmingen). Antibodies used for Western blot were anti-phospho Erk1/2 mouse monoclonal antibody, Erk1/Erk2 rabbit polyclonal antibody, anti–phospho Akt (protein kinase B; Ser473) rabbit antibody and anti–Akt rabbit polyclonal antibody (Cell Signaling Tech, Beverly, MA). The secondary antibodies were peroxidase-labeled anti–mouse immunoglobulin G (IgG) and anti–rabbit IgG horseradish peroxidase (HRP)–linked antibodies (Amersham Biosciences, Piscataway, NJ). Mitogen-activated protein/extracellular signal regulated kinase (MEK) inhibitor, PD98059, was purchased from Cell Signaling. TGF-β1 was purchased from R&D (R&D Systems, Minneapolis, MN) and resuspended and stored as per the manufacturer’s instructions. Human recombinant SDF-1 was purchased from BioVision (Mountain View, CA).

**Cells**

Human cord blood was obtained with institutional review board approval. Low-density mononuclear cells were isolated by density gradient centrifugation over Ficoll-Paque Plus (Amersham Pharmacia Biotech). CD34+ cells were enriched using direct CD34 magnetic separation kit (Miltenyi Biotech) according to the manufacturer’s instructions. Purity of the enriched CD34+ cells was 95% to 98% after cells were sequentially passed over 2 columns.

**Expansion and pretreatment of CD34+ cord blood (CB) cells**

Enriched CD34+ cells were expanded by growing them in Iscoves modified Dulbecco medium (IMDM; Gibco, Grand Island, NY) containing 10% fetal calf serum (FCS), L-glutamine, and including 100 ng/mL recombinant human (rh) thrombopoietin (TPO), 100 ng/mL rhFlt-3 ligand, and 50 ng/mL rh stem cell factor (SCF) for 4 days at 37°C in a humidified atmosphere containing 5% CO2. After 4 days, cells were centrifuged, and the pellet was resuspended in medium containing the above-mentioned cytokines. Cells were pretreated in medium (containing SCF, TPO, Flt-3 ligand) alone or in medium (containing SCF, TPO, and Flt-3 ligand) along with SDF-1 (200 ng/mL) with or without TGF-β1 (0.5 ng/mL) for 24 hours, unless otherwise indicated. All pretreatments contained SCF, TPO, and Flt-3 ligand at the above-mentioned doses. After incubation, cells were harvested, washed, and used in different assays. In some experiments, freshly isolated CD34+ cells were cultured in IMDM containing 2 mM L-glutamine and 20% FCS alone or containing SDF-1 (200 ng/mL) with or without TGF-β1 (0.5 ng/mL) for 20 to 24 hours and used in chemotaxis and adhesion assays.

**Migration assay**

Chemotaxis assays were performed using 96-well chemotaxis chambers (NeuroProbe, Gaithersburg, MD) in accordance with the manufacturer’s instructions with minor variations. CD34+ cells in 30 µL chemotaxis medium (IMDM + 1% FCS) were added to the membrane. Chemotaxis medium alone (300 µL) or containing 200 ng/mL SDF-1 was added to the bottom well. After 4 hours of incubation at 37°C in 5% CO2, cells migrated to the lower chamber, and nonmigrated cells in the upper chamber were counted for 30 seconds using fluorescence-activated cell scan (FACScan) under identical flow conditions. In experiments designed to determine migration of progenitor cells to SDF-1, both input and migrated cells were counted using a Coulter counter prior to plating cells in semisolid culture.

**Progenitor cell colony assay**

To assess clonogenic ability, methylcellulose progenitor assay was performed as described previously. Briefly, 1 to 5 x 10^2 CD34+ cells/mL were plated in 1% methyl cellulose, 10^{-4} M 2-mercaptoethanol, 2 mM L-glutamine, 30% fetal calf serum, 1 U/mL recombinant human (rh) erythropoietin, 10 ng/mL rh granulocyte-macrophage colony-stimulating factor (GM-CSF), 10 ng/mL rh interleukin-3, and 50 ng/mL rhSCF. Cultures, set up in duplicate or triplicate, were incubated in a humidified atmosphere, 5% CO2 and 5% O2 at 37°C for 14 days. Colonies from erythroid burst colony-forming units (BFU-Es), and mixed erythroid-myeloid colony-forming units (CFU-GEMMs) were scored.

**Flow cytometry analysis of cell surface marker**

Freshly enriched CD34+ cells or cytokine-expanded CD34+ cells were washed, resuspended in FACSScan buffer (PBS (phosphate-buffered saline) + 2 mM EDTA (ethylenediaminetetraacetic acid) + 2% FCS), incubated for 5 minutes at room temperature with Fe-block, and then incubated with a recommended volume of anti-human antibodies: CD34 conjugated to FITC, CXCR4 conjugated to PE and CD38 conjugated to APC. Cells were incubated for 30 minutes at 4°C, washed, and fixed in 2% paraformaldehyde prior to analysis by FACSscan using CellQuest software (BD Biosciences Immunocytometry Systems, San Jose, CA).

**Adhesion assay**

Adhesion assays were performed in 96-well plates (high-binding; Costar, Cambridge, MA) coated with purified plasma fibronectin (Sigma, St Louis, MO; 10 µg/mL) overnight at 4°C in 100 mM bicarbonate buffer (pH 8.8), followed by 2 hours of incubation at 37°C. To block nonspecific binding sites, plates were subsequently incubated for another 2 hours at 37°C with 2% bovine serum albumin (BSA) in the same solution. CD34+ cells were added to wells and incubated at 37°C in a humidified atmosphere containing 5% CO2 for 30 minutes. To distinguish between transient and sustained adhesion mediated by SDF-1, SDF-1 was added to the wells either in the last 2 minutes of incubation (transient response) or at the start of the incubation and kept there until the end of the incubation period (sustained adhesion). In certain experiments, cells were preincubated with either 10 µg/mL human integrin-specific antibody or isotype control antibody at 4°C for 30 minutes or with PD98059 (40 µM) at 37°C for 30 minutes. Cells were washed and plated for adhesion assay. Antibodies did not contain azide. After incubation, nonadherent cells were removed, and the wells were washed 2 to 3 times with medium. Adherent cells were recovered by detaching them using Cell Dissociation Buffer (Gibco).

**Actin polymerization**

Actin polymerization assays were performed as described by others with a few modifications. Cells pretreated with different stimuli were washed and resuspended in IMDM supplemented with 0.1% BSA at 10^5 cells/mL. SDF-1 (200 ng/mL) was added to the cell suspension. At various time intervals, cells were permeabilized and fixed using 0.2 mL permeabilizing/fixed solution (BD Cytofix/Cytoperm; BD Biosciences Pharmingen) and stained with 0.1 mL Phalloidin-FITC solution (4 x 10^{-7} M; Sigma). Cells were incubated for 10 minutes at 37°C, and washed twice with BD wash buffer (BD Biosciences Pharmingen, Palo Alto, CA). Pellets were resuspended in 500 µL 2% paraformaldehyde solution. Mean fluorescence was measured by FACScan.

**Calcium influx**

Calcium flux induced by SDF-1 in CD34+ cells was studied by flow cytometry. An equal volume of Fluo-3/AM (stock concentration 2 mM; Molecular Probes, Eugene, OR) and Fluronic acid (stock concentration 20% wt/vol; Molecular Probes) were mixed just before use. Cells were washed and resuspended in IMDM + 2% BSA, and Fluo-3/AM/Phuronic acid mix was added for a final Fluo-3/AM concentration of 4 µM. After incubation for 45 minutes at room temperature, cells were washed in Ca2+ flux assay buffer (Hanks Balanced Salt solution containing 20 mM HEPES (N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid) and 0.2% BSA, pH 7.4) to remove extracellular dye, incubated for 10 minutes at room temperature, and analyzed by FACScan. Background fluorescence of each sample was measured, and SDF-1 was then added to samples. Samples were quickly mixed by vortex, and Ca2+ influx was recorded.

**Western blot**

Cells pretreated with different stimuli were washed, resuspended in IMDM + 0.5% BSA, and either left unstimulated or stimulated with SDF-1 for the indicated time period. Alternatively, cells were first incubated with PD98059 for 45 minutes at 37°C before SDF-1 stimulation. Cells were immediately centrifuged at 4°C and washed with ice-cold PBS prior to lysis, followed by centrifugation at 14 000g for 30 minutes at 4°C.
content of lysates was quantified using bicinchoninic acid protein assay reagent (Pierce, Rockford, IL), and samples were adjusted to equal protein concentration and volume. Total cell lysates were resolved in 10% sodium dodecyl sulfate (SDS)–polyacrylamide gels (Novex, San Diego, CA) and transferred to Hybond membrane (Millipore, Bedford, MA). Filters were blocked using 3% BSA in Tris (tris(hydroxymethyl)aminomethane)–buffered saline/Tween-20 (TBST) for 1 hour and incubated overnight with antibodies. Membranes were washed with TBST and incubated with secondary antibodies conjugated to horseradish peroxidase, and antibody binding was detected by enhanced chemiluminescence (ECL) reaction (Amersham Life Science, Arlington Heights, IL).

Statistical analysis

Student 2-tailed t test was used for statistical analysis. The level of significance is indicated by P value.

Results

Effect of in vitro expansion of CD34+ CB cells on CXCR4 and CD34 expression

Earlier studies demonstrated that the cocktail of TPO, Flt-3 ligand, and SCF induces increased numbers of CD34+ and hematopoietic progenitor cells from human CD34+ cells. Expanded progenitor cells using the above-mentioned cytokines retained the ability to engraft in vivo. In this study we have used cytokine-expanded CD34+ cells, and selected experiments were performed using freshly isolated CD34+ cells. We expanded freshly isolated CD34+ CB cells for 4 days using the combination of TPO, Flt-3 ligand, and SCF. After 4 days of expansion, there was an approximately 3.5-fold increase in the total number of cells. Moreover, the cells still remained essentially CD34+ positive, albeit levels of CD34 and CXCR4 on the expanded population were more heterogeneous than freshly enriched CD34+ cells (Figure 1A). Expanded CD34+ cells were then treated with the same cytokine cocktail (TPO, Flt-3, SCF) either alone or containing TGF-β1, SDF-1, or SDF-1 and TGF-β1 for 24 hours. SDF-1 pretreatment decreased the proportion of cells expressing CXCR4 and also decreased CXCR4 expression (Figure 1B). However, SDF-1 along with TGF-β1 pretreatment increased the proportion of cells expressing CXCR4 but did not change CXCR4 expression level (Figure 1B).

Desensitization of CD34+ cells to SDF-1

Prolonged exposure of cells to SDF-1 induces desensitization and internalization of CXCR4, leading to reduced SDF-1–induced chemotaxis. Similarly, we found that calcium mobilization in response to SDF-1 was significantly desensitized in freshly isolated CD34+ cells by prior treatment with SDF-1, either for 1 hour or 16 hours (Figure 2). Further to determine how long this desensitization lasts, SDF-1–pretreated CD34+ cells were kept in medium alone for varying lengths of time, and then calcium mobilization in response to SDF-1 was analyzed. As seen in Figure 2, SDF-1 withdrawal for 4 hours, but not 1 hour, partially restored SDF-1–induced calcium mobilization in SDF-1–pretreated CD34+ cells.

Figure 2. Desensitization of CXCR4. Nonexpanded CD34+ cells were pretreated with SDF-1 (200 ng/mL) for 1 hour, 20 hours, or kept in medium alone. Cells were washed and either immediately loaded with Fluo-3AM or kept in medium for 1 hour or 4 hours (in the absence of SDF-1) and then loaded with Fluo-3AM. The time noted in parentheses in the insert key box denotes the time for which SDF-1–pretreated cells were kept in medium prior to SDF-1 stimulation. Baseline fluorescence was first recorded and calcium flux following SDF-1 (200 ng/mL) stimulation (indicated by ↑) was analyzed by FACScan. Increase in fluorescence intensity following SDF-1 addition (compared with baseline), represents the calcium flux in cells and is shown on the y-axis of the plot. Data represent mean ± SEM of 3 independent experiments.* P < .05 and **P < .05 compared with cells pretreated in with SDF-1 for 1 hour and 20 hours, respectively.
**TGF-β1 modulates chemotactic response of CD34+ cells to SDF-1**

Considering the importance of SDF-1/CXCR4 interactions in vivo, we investigated the chemotactic response of ex vivo-expanded CD34+ cells, prestimulated with SDF-1 for 24 hours, to SDF-1 in an in vitro chemotaxis assay. SDF-1 pretreatment reduced the chemotactic response of CD34+ CB cells to SDF-1 dose dependently (Figure 3A). Down-modulation in chemotactic response of the SDF-1–prestimulated CD34+ cells to SDF-1 was significantly overcome when the cells were prestimulated with SDF-1 in the presence of TGF-β1 (Figure 3B). The effect of TGF-β1 was dose dependent; 0.5 ng/mL TGF-β1 was optimal (Figure 3C). Reduction in SDF-1–induced migratory response in SDF-1–pretreated cells could be seen as early as 2 hours and lasted up to 24 hours of SDF-1 pretreatment (Figure 3D). To determine whether the effect of TGF-β1 was time dependent, CD34+ cells were coincubated with TGF-β1 and SDF-1 for varying lengths of time up to 24 hours before being subjected to SDF-1–induced chemotaxis. Maximal restoration of SDF-1–induced chemotactic response was observed when cells were cultured with TGF-β1 and SDF-1 for 24 hours (Figure 3D). Shorter coincubation of cells with TGF-β1 and SDF-1 restored partially, but not significantly, SDF-1–induced chemotactic response.

To explore whether this phenomenon was observed in fresh CD34+ cells, we pretreated nonexpanded CD34+ cells with SDF-1 alone or SDF-1 and TGF-β1 for 20 hours and compared their chemotactic activity toward SDF-1. As shown in Figure 3E, similar to cytokine-expanded CD34+ cells, SDF-1–pretreated cells had reduced chemotactic activity compared with CD34+ cells cultured in medium alone, and this was partially restored by TGF-β1.

**Analysis of migrated cells–cell surface maker and clonogenic potential**

We next examined whether this effect was reproduced in a more primitive hematopoietic cell compartment. CD34+/CD38lo cells identify primitive hematopoietic populations. Similar to total CD34+ population, SDF-1–directed migratory response of CD34+/CD38lo cells was reduced by SDF-1 pretreatment (Figure 4). This effect was observed for both cytokine-expanded (Figure 4A) and freshly isolated cells (Figure 4B). Reduction in migration potential of CD34+/CD38lo cells was significantly reversed when these cells were exposed simultaneously to SDF-1 and TGF-β1. SDF-1–induced chemotactic response of erythroid and multipotential progenitors was significantly reduced by SDF-1 pretreatment but was restored when CD34+ cells were preincubated with SDF-1 in the presence of TGF-β1 (Figure 4C-D).

**Actin polymerization and calcium flux**

Transient actin polymerization in response to chemokines is an important cellular event for cell polarization, formation of cell membrane structures such as uropods, and chemotaxis. Therefore, we examined whether SDF-1–induced actin polymerization (F-actin) was affected by preexposure of CD34+ cells to SDF-1. SDF-1 pretreatment led to a significant reduction in SDF-1–induced actin polymerization (F-actin) in both ex vivo cytokine-expanded (Figure 5A) and freshly isolated (nonexpanded) CD34+ cells (Figure 5B) prestimulated with SDF-1, compared with cells cultured in SDF-1 and TGF-β1. Although the amount of SDF-1–induced actin polymerization in CD34+ cells pretreated with SDF-1 and TGF-β1 was reproducibly and significantly higher than that induced in CD34+ cells pretreated with SDF-1 alone, it was lower than SDF-1–induced actin polymerization in cells cultured in medium alone (Figure 5A-B).

Consistent with a previous report, we found that the CXCR4-mediated rise in cytosolic free calcium is desensitized by prior exposure of cells to SDF-1. This activity was only partially restored when cells were exposed to SDF-1 in the presence of TGF-β1 (Figure 5C expanded) and D (nonexpanded).
basal level of Erk1/Erk2 phosphorylation was comparable among the cells pretreated under different conditions (Figure 6A). CD34+ cells pretreated with cytokine cocktail alone or along with TGF-β1 displayed high levels of SDF-1–induced Erk1/Erk2 phosphorylation, whereas cells pretreated with SDF-1 failed to do so. When CD34+ cells were pretreated with SDF-1 in the presence of TGF-β1, they retained the ability to undergo robust Erk1/Erk2 phosphorylation in response to SDF-1 stimulation.

Unlike SDF-1–induced Erk1/Erk2 phosphorylation, SDF-1 did not induce any Akt phosphorylation above the background level in cells pretreated under different conditions (Figure 6A). Moreover background Akt phosphorylation levels were comparable among cells pretreated with cytokine cocktail alone or cytokine cocktail and SDF-1 alone, TGF-β1 alone, or SDF-1 along with TGF-β1.

To examine the requirement for Erk activation in SDF-1–induced migration, the migration assays were performed in the presence of PD98059, a MEK inhibitor. PD98059-inhibited migration potential of CD34+ cells cultured in cytokine cocktail alone or pretreated with TGF-β1 or with SDF-1 and TGF-β1 was significantly reduced and was comparable to cells pretreated with SDF-1 alone (data not shown). We consistently observed that the migration potential of cells pretreated with SDF-1 and TGF-β1 was significantly inhibited in the presence of PD98059, whereas PD98059 did not have much effect on the residual migration potential of SDF-1–pretreated cells (Figure 6B). This suggests that phosphorylation of Erk1/Erk2 is required for maximal SDF-1–induced chemotaxis. Residual chemotactic activity seen in CD34+ cells pretreated with SDF-1 was unaffected by Erk1/Erk2 phosphorylation and possibly involves other signaling pathways.

Short-term and long-term adhesion mediated by SDF-1

HSPCs in the bone marrow (BM) are thought to be positioned within specific stromal niches that provide the microenvironment for their proliferation and differentiation.44 As HSPCs differentiate, they move from one type of niche to another.45,46 For HSPCs to remain in bone marrow for prolonged periods, a sustained adhesive response needs to be triggered. We examined the effect of SDF-1 preexposure on both SDF-1–induced transient (short-term, 2-minute SDF-1 stimulation) and sustained adhesion (long-term, 30-minute SDF-1 stimulation) responses. In expanded CD34+ cells, SDF-1 induced a significant increase in transient adhesion response, irrespective of pretreatment conditions. This response was not affected by SDF-1 preexposure (Figure 7A). Pretreatment of cells with TGF-β1 along with SDF-1 did not further enhance this response. Next, to simulate the bone marrow situation whereby the stem/progenitor cells are continuously exposed to SDF-1, we evaluated the effect of long-term SDF-1 stimulation of cytokine-expanded CD34+ cells on their adhesive response. We found that the sustained adhesion induced by long-term SDF-1 stimulation was reduced by prior exposure of the cells to SDF-1, and it could be significantly recovered if the cells were pretreated with SDF-1 in the presence of TGF-β1. To examine whether the SDF-1–induced sustained adhesion was dependent on VLA-4/VLA-5 activation, cells were incubated with either anti–VLA-4 or anti–VLA-5 antibodies before the adhesion assay. Irrespective of the pretreatment conditions, we found that SDF-1–induced sustained adhesion could be significantly inhibited by both anti–VLA-4 and anti–VLA-5 antibodies (Figure 7B). We also evaluated the role of Erk, an important molecule in SDF-1 and integrin-triggered signaling pathways.42–44 We used PD98059, a MEK inhibitor, at a concentration of 40 μM. PD98059 significantly inhibited SDF-1–induced sustained adhesion by 59.37%, 43.57%, 60.4%, and 57.3% (P < .05)

SDF-1–induced Erk1/Erk2 but not protein kinase B (PKB)/Akt phosphorylation is down-modulated by SDF-1 pretreatment

Erk-mitogen kinase pathway has been implicated in cell migration39–41 and can be activated by both SDF-142 and TGF-β1.43 Therefore, we examined the levels of SDF-1–induced Erk1/Erk2 phosphorylation in cells pretreated under various conditions. The
for cells pretreated with cytokine cocktail alone or along with TGF-β1, SDF-1, or SDF-1 and TGF-β1, respectively.

In nonexpanded CD34+ cells, SDF-1 induced a significant sustained adhesion response irrespective of pretreatment conditions; however, the greatest effect was observed when cells were pretreated with SDF-1 and TGF-β1 (Figure 7C). Short-time exposure (2 minutes) of nonexpanded CD34+ cells to SDF-1 induced only marginal increases in adhesion response above the basal response. The SDF-1–induced transient adhesion response of CD34+ cells could be significantly blocked by anti-VLA-4 antibody (Figure 7D).

Discussion

Stem cells are localized in a microenvironment known as the stem cell niche, where they are maintained in an undifferentiated and quiescent state. During development of blood cells, stem and progenitor cells translocate from a quiescent into a permissive proliferative vascular niche.48,49 Several studies in mice have demonstrated that both the SDF-1/CXCR4 and VLA4/VCAM-1 (vascular cell adhesion molecule-1) axes are critical for hematopoiesis and retention of hematopoietic stem/progenitor cells in bone marrow.50,51 SDF-1 transiently activates cell surface integrins in CD34+ bone marrow progenitor cells.52,53 Although transient activation of SDF-1–mediated adhesion may explain the homing of progenitor cells to bone marrow and also trafficking of blood cells to inflammation sites,52,54,55 it does not explain the scenario in bone marrow microenvironment where stem/progenitor cells are retained for prolonged periods. In bone marrow, stem and developing progenitor cells need to be retained in appropriate niches for survival, proliferation, and/or differentiation.49 Unlike the peripheral vasculature, there is constitutive high expression of VCAM-1 and SDF-1 in bone marrow.53,55 These findings suggest that in bone marrow, although cells are exposed to high concentrations of SDF-1, they remain responsive to SDF-1 and are not only retained in the appropriate niches, but, when required, they can move from one niche to the other and finally exit into the peripheral blood.57 Paradoxically, it has been shown that exposure of cells to SDF-1 makes them unresponsive to SDF-1 because of CXCR4 receptor desensitization.53 Given these opposing findings, it appeared that in bone marrow, responsiveness of the stem/progenitor cells to SDF-1 must be modulated by other factors. In this study we describe TGF-β1 as a modulator of long-term responsiveness of CD34+ stem/progenitor cells to SDF-1. In the presence of TGF-β1, CD34+ cells preexposed to SDF-1 remain chemotactically responsive to SDF-1 for a prolonged period of time and retain sustained adhesion response mediated by SDF-1. Consistent with earlier reports33,38,58 we found that calcium mobilization in response to SDF-1 was significantly dampened by prior exposure of CD34+ cells to SDF-1. Furthermore when CD34+ cells are exposed to SDF-1 for 20 to 24 hours, their migration response to SDF-1 is significantly reduced. This phenomenon was observed for both freshly isolated (nonexpanded) and cytokine-expanded CD34+ cells. However, TGF-β1 significantly restored chemotactic response in both freshly isolated and cytokine-expanded SDF-1 preexposed CD34+ cells.

Figure 6. Improved chemotactic activity of cells pretreated with SDF-1 and TGF-β1 is associated with enhanced Erk1/Erk2 phosphorylation in response to SDF-1. (A) Ex vivo–expanded CD34+ cells were cultured in cytokine cocktail alone (medium) or along with TGF-β1, SDF-1, or SDF-1 and TGF-β1 for 24 hours. Cells were harvested, washed, and either left unstimulated or stimulated with 200 ng/mL SDF-1 for 3 minutes. Cells were solubilized, and extracts were subjected to immunoblotting by using anti–phospho Erk1/Erk2 or anti–phospho-Akt (Ser473) antibodies. After stripping and saturating nonspecific protein binding, we reprobed the same blot with anti–Erk1/Erk2 or anti–Akt antibody. Blots were developed by a chemiluminescence reaction and exposed to radiographic film. A representative experiment of 1 of 3 experiments performed is shown. (B) Effect of MEK inhibitor, PD98059, on migration activity of CD34+ cells pretreated with cytokine cocktail along with SDF-1 (200 ng/mL) or SDF-1 (200 ng/mL) + TGF-β1 (0.5 ng/mL) SDF-1 for 24 hours. Cells were tested for their migration response to SDF-1 (200 ng/mL). Data represent the mean ± SD of 4 independent experiments.
Figure 7. SDF-1 induces adhesion of CD34+ cells to fibronectin. (A) Ex vivo–expanded CD34+ cells were pretreated with cytokine cocktail alone (medium) or containing TGF-β1 (0.5 ng/mL), SDF-1 (200 ng/mL), or SDF-1 (200 ng/mL) + TGF-β1 (0.5 ng/mL) for 24 hours. The cells were washed and then plated on fibronectin (10 μg/mL) coated wells and incubated for 30 minutes. Cells were either left unstimulated (□) or were stimulated with SDF-1 (200 ng/mL) for the full 30 minutes (▲) or only for the last 2 minutes (■) of the adhesion assay. Data represent the mean ± SD of 3 independent experiments. *P < .05 compared with adhesion response in the absence of SDF-1 stimulation (basal adhesion) for respective pretreatment. (B) Ex vivo–expanded CD34+ cells pretreated with SDF-1 alone, CD34+ cells pretreated under various conditions were preincubated with anti–VLA-4 antibody and assayed in adhesion assay. The cells were plated in fibronectin-coated plates and stimulated with SDF-1 for 30 minutes. Data represent the mean ± SD of 3 independent experiments. *P < .05 compared with adhesion response in the absence of antibodies. (C) Nonexpanded CD34+–enriched cord blood cells were cultured in medium (containing 20% FCS) alone or along with TGF-β1 (0.5 ng/mL), SDF-1 (200 ng/mL), or SDF-1 (200 ng/mL) and TGF-β1 (0.5 ng/mL) for 24 hours. These cells were then assayed for their ability to bind to fibronectin in the absence or presence of 200 ng/mL SDF-1 for 2 minutes (transient response) and 30 minutes (sustained response). Nonadherent cells were removed, and adherent cells were quantified as described in "Materials and methods." Data represent the mean ± SEM of 3 independent experiments. *P < .05 compared with adhesion response in absence of SDF-1 stimulation (basal adhesion) for respective pretreatment. (D) Fresh CD34+ cells pretreated under various conditions for 24 hours were preincubated with anti–VLA-4 antibody and assayed in adhesion assay as described in "Materials and methods." Data represent mean ± SEM of 1 experiment. *P < .05 compared with adhesion response in absence of antibodies.

CXC4 signaling is involved in the regulation of integrin function.15,53 We found that SDF-1 induced both sustained and transient adhesion responses to fibronectin in cytokine-expanded CD34+ cells. The SDF-1–induced sustained adhesion response was impaired by SDF-1 preexposure but near normal if cells were pretreated with SDF-1 in the presence of TGF-β1. This effect was seen for both expanded and nonexpanded CD34 cells. Unlike sustained adhesion, SDF-1–mediated transient adhesion response in CD34+ cells remained unaffected by SDF-1 preexposure. These findings suggest that not all responses in CD34+ cells mediated by SDF-1 are attenuated by preexposure to SDF-1. Since SDF-1 activates different signaling pathways, it is possible that some pathways are more sensitive to receptor desensitization than the others and/or are likely to be regulated at different levels. An earlier study showed that in B cells, although both SDF-1–mediated transient and sustained adhesion involved focal adhesion kinase (FAK) phosphorylation, upstream signaling pathways were qualitatively different.67

It has been recently reported that TGF-β1 down-regulates expression of SDF-1 by stromal cells, resulting in reduced cell migration and adhesion of BM cells, and it was suggested that this could affect the motility of cells in the bone marrow.68 A concentration of 2 ng/mL TGF-β1 was used in that study. We found that 0.5 ng/mL TGF-β1 was optimal for improving SDF-1–mediated responses in CD34+ cells (preexposed to SDF-1). Increasing the dose of TGF-β1 not only resulted in reduced migration but also led to apoptosis (data not shown) in these cells. In physiologic settings the level of active TGF-β1 is low and highly regulated.69 However, in several pathologic conditions it is overproduced and leads to...
failure of early hematopoietic progenitors.\textsuperscript{70,71} It is, therefore, likely that TGF-\(\beta\)1 modulates SDF-1-mediated responses, but the outcome depends on the dose of TGF-\(\beta\). The dose-dependent effect of TGF-\(\beta\)1 has also been seen in its ability to support myeloid cell proliferation.\textsuperscript{72}

In summary, we show that both SDF-1–stimulated chemotactic activity and sustained adhesion of CD34\(^{+}\) cells is down-regulated by preexposure of CD34\(^{+}\) cells to SDF-1, and these responses are modulated by TGF-\(\beta\)1. Since SDF-1 is constitutively present at high concentrations in the bone marrow compartment, modulation of SDF-1 responses by TGF-\(\beta\)1 may explain, at least in part, how hematopoietic cells remain responsive to SDF-1 in bone marrow. Therefore, modulation of SDF-1 activity by TGF-\(\beta\)1 may be an essential physiologic process that plays a role in retention of stem/progenitor cells in their niches and their movement among the niches.

References


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