Brief report

Phenotypic characterization of the human myeloma cell growth fraction

Nelly Robillard, Catherine Pellat-Deceunynck, and Régis Bataille

In this study we quantified the proliferation rate of normal and malignant plasma cells (PCs) by ex vivo incorporation of 5-bromo-2’-deoxyuridine (BrdU; labeling index, LI) using flow cytometry. We show that all bone marrow PCs, either normal or malignant, include a subset of proliferating PCs present within the CD45bright fraction. Indeed, medullary normal and malignant PCs were always heterogeneous for CD45 expression, and proliferation was always restricted primarily to the CD45bright compartment. Moreover, an inverse correlation was found between LI or CD45 and B-cell lymphoma 2 (Bcl-2) in both malignant and normal PCs, the most proliferating CD45bright PCs have the lowest Bcl-2 expression. We investigated expression of molecules of interest in multiple myeloma (MM)—that is, CD138, CD19, CD20, CD27, CD28, CD56, and CD11a—to further characterize the CD45bright fraction. Among all of these molecules, only CD11a was exclusively expressed by CD45bright proliferating myeloma cells. In conclusion, proliferating myeloma cells are characterized by the specific CD45bright CD11apos Bcl-2low phenotype. (Blood. 2005;105:4845-4848)

Introduction

Multiple myeloma (MM) is primarily conceptualized as an accumulative disease. Indeed, most studies evaluating the labeling index (LI) of plasma cells (PCs) revealed that it rarely exceeded 1%. However, as early as 1981, Drewinko et al1 showed that in vivo (LI) of plasma cells (PCs) revealed that it rarely exceeded 1%. However, as early as 1981, Drewinko et al1 showed that in vivo proliferation of myeloma cells was generally nonproliferating, although a small fraction of them could proliferate (growth fraction, GF). Thus, they proposed the concept that this minor GF could give rise to the major nonproliferating fraction.

Myeloma cells either lack or express a weak to intermediate level of CD45.2,3 However, in previous studies we have shown that CD45 and also CD11a are frequently expressed by a subset of myeloma cells only.2 It has been demonstrated that CD45 expression is highly correlated with the proliferation rate of myeloma cells.4,5 With regard to normal counterparts, PCs are heterogeneous in terms of CD45 phenotype. Recently, Medina et al6 confirmed the association of maturity with decreasing CD45 expression.7-9 Generation of PCs from B cells, mainly studied in vitro in humans, is a multistep process that involves both proliferation and maturation/differentiation.10-13 The aim of this study was to evaluate the proliferation of different types of normal PCs in relation to their phenotype, especially to CD45, to understand the biology of polyclonal and monoclonal PC expansions that are reactive plasmacytoses (RP) and MM, respectively.

Study design

Samples and reagents

Forty-nine consecutive patients with MM (25 at diagnosis, 24 at relapse) were included in this study. Bone marrow and blood samples from healthy donors or patients with reactive plasmacytosis, and tonsil samples were obtained and prepared as described.1,11 Approval was obtained from the Nantes University Hospital Institutional Review Board for these studies. Informed consent was provided according to the Declaration of Helsinki. Antibodies directed against (1) CD11a, CD19, CD45, CD138, Apo2.7 and (2) CD28, CD38, CD56, 5-bromo-2’-deoxyuridine (BrdU) were from Beckman Coulter (Miami, FL) and BD Biosciences (San Jose, CA), respectively.

Cell staining

Mononuclear cells (MNCs; 0.5-3 × 10⁶) were stained in a 4-color assay with anti-CD45–fluorescein isothiocyanate (FITC; J33), anti-CD138–phycoerythrin cyanine 5 (PECy5; B-B4), anti-CD38–APC (HB7) and control-PE, anti-CD56–PE (MY31), anti-CD28–allophycocyanin (PE; L293), anti-CD19–PE (J4.119), anti-APO2.7–PE (2.7A6A3), and anti-CD11a–PE (25.3) monoclonal antibodies (mAbs) as described.14 To determine CD45-FITC fluorescent staining, MNCs were stained with isotype-FITC, anti-CD38–APC, anti-CD45–PECy5, and anti-CD138–PE mAbs. For intracellular Bcl-2 staining, MNCs were first stained with anti-CD38–APC, anti-CD45–PECy5, and anti-CD138–PE mAbs, then with anti-Bcl-2–FITC (124; Dako, Glostrup, Denmark) or control isotype-FITC mAbs after permeabilization (Intra Prep; Beckman Coulter). For LI, cells were incubated overnight with or without 50 μM BrdU (5-bromo-2’-deoxyuridine; Sigma, St Louis, MO) at 37°C in culture medium, then stained with anti-CD45–APC, anti-CD38–PE, and anti-CD138–PECy5 mAbs (with anti-CD45–APC and anti-CD38–PE mAbs for tonsil and blood PCs), permeabilized, and stained with anti-BrdU as described.12

Flow cytometry analysis

Data were acquired by means of a Becton Dickinson fluorescence-activated cell sorting FACSCalibur with CellQuest Pro software (BD Biosciences).14 PCs were identified using a sequential gating strategy (Figure 1).
Myeloma cells and bone marrow PCs were heterogeneous: proliferation was restricted to the CD45bright SSC^high compartment

The percentage of BrdU-positive myeloma cells was low, generally around 1% (n = 49 patients). In all patients, myeloma cells were heterogeneous for CD45 expression and SSC, allowing a definition of 2 compartments: CD45^{bright}SSC^{high} and CD45^{int/low}SSC^{low}. The CD45^{bright}SSC^{high} compartment was always present, representing 12% (median) of myeloma cells (illustrated in Figure 1). The second compartment CD45^{int/low}SSC^{low} represented the majority of tumor cells (88%). We observed that the LI of the CD45^{bright}SSC^{high} compartment was always greater than that of the CD45^{int/low}SSC^{low} compartment and far above 1% (median, 6.8%; n = 49; Table 1). In general, the CD45^{bright}SSC^{high} compartment had a 6.8 higher proliferative level than the CD45^{int/low}SSC^{low} compartment (P < .01). As outlined in Table 1, the LI of the CD45^{bright} compartment could reach 40% of myeloma cells, which was similar to what we observed in reactive plasmacytoses, suggesting that in these patients, almost all CD45^{bright} myeloma cells were proliferating. Of note, the total LI correlated highly with the LI of the CD45^{bright} compartment (r = 0.81, P < .001) and less with that of the CD45^{low} compartment (r = 0.44, P < .01).

As observed for myeloma cells, 2 compartments of PCs delineated by CD45 and SSC were found in normal bone marrows (n = 11, illustrated in Figure 1). The CD45^{bright}SSC^{high} compartment, which represented 65% of the PCs, was highly proliferative (LI = 18.4% for BM1). The second CD45^{int/low}SSC^{low} compartment included 35% of the PCs with a much lower LI: 2.6% (BM1 and Table 1). A subset of PCs which lacked CD45 expression was observed in 2 cases.

**Figure 1.** CD45 phenotype and LI of myeloma and bone marrow PCs. PC phenotype analysis was performed in 2 steps. In the first step, 15,000 total events were acquired to draw a PC gate in the side scatter (SSC)/CD38^{+} dot-plot (R1). In the second step, an acquisition of 15,000 PCs (at least 1000 PCs for very low plasmacytoses) was performed through the R1 live-gate. PCs were identified by coexpression of CD38 and CD138 (gate R2). A third region was set on the light scatter of the cells satisfying both R1 and R2 to exclude debris or apoptotic PCs with a low CD138 expression and a characteristic light-scatter distribution.12 The lack of Apo 2.7 staining in the gated population confirmed that cells were not apoptotic.13 Then 2 PC subpopulations were identified on the CD45 versus SSC dot plot: R3 was set around PC with a large SSC expressing a high level of CD45, while R4 was set around PC with low SSC and negative or weak CD45 expression. The analyses of the phenotype and the LI were performed in these 2 PC subpopulations, separately and simultaneously. The percentage of BrdU^{+} PCs within the population incubated with BrdU (BrdU^{+}) or without (control, BrdU^-) were indicated within the cytograms. For example, in patient MM1 the global LI was 1.7% (3.1%–1.4%) but 8.5% (9.1%–0.6%) in R3 and 1.4% (1.5%–0.1%) in R4. Overlay histograms represent the immunofluorescence of Bcl-2 (thick line) over the control (thin line). r indicates the ratio of MFI.

**Statistical analysis**

Statistical analyses were performed using nonparametric Wilcoxon rank sum, Spearman correlation coefficient, and the sign tests.

**Results and discussion**

**Myeloma cells and bone marrow PCs were heterogeneous: proliferation was restricted to the CD45^{bright} SSC^{high} compartment**

Since we have previously observed that Bcl-2 inversely correlates with the LI in both reactive PCs and myeloma cells, we looked for a correlation within myeloma and PC subsets (Figures 1-2). We found that CD45^{bright} myeloma cells always expressed a lower level of Bcl-2 compared with that of CD45^{low/bright} myeloma cells (P < .01). In normal bone marrow (BM), a significant increase in Bcl-2 MFIR was found that in these patients, almost all CD45^{bright} myeloma cells were proliferating. Of note, the total LI correlated highly with the LI of the CD45^{bright} compartment (r = 0.81, P < .001) and less with that of the CD45^{low} compartment (r = 0.44, P < .01).

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**BrdU^{+} myeloma cells expressed lower Bcl-2 levels**

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**Phenotypic characterization of the CD45^{bright} SSC^{high} proliferating compartment in MM: only CD11a expression was restricted to the CD45^{bright} SSC^{high} subset**

We further characterized the phenotype of the CD45^{bright} myeloma cells using relevant MM markers (CD19, CD20, CD27, CD28, and CD56). As illustrated in Figure 3, CD19, CD27, CD28, and CD56 were expressed or not by myeloma cells, and their expression was unrelated to CD45 subsets. However, we found that CD11a
overlapped completely with the CD45bright compartment in all the patients studied (Figure 3), while CD45low/neg myeloma cells were negative for CD11a. Ahsmann et al. have published that lymphocyte function-associated antigen 1 (LFA-1) (CD11a-CD18) expression correlated with tumor growth in MM. LFA-1 is involved in either homotypic or heterotypic interactions in MM. Indeed, human myeloma cells, like stromal cells, express the 3 ligands of LFA-1: intercellular adhesion molecule-1 (ICAM-1) (CD54), ICAM-2 (CD102), and ICAM-3 (CD50). These interactions, restricted to

Table 1. CD45 expression and labeling index of PCs

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<th>Cell type</th>
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MFIR indicates mean of fluorescence intensity ratio; PCLI, plasma cell labeling index; and —, no data (no cells).

Figure 2. CD45 phenotype and LI of tonsillar, peripheral normal, and reactive PCs. Analysis of phenotype, LI, and Bcl-2 expression in PCs isolated from tonsil and blood. PCs were identified by CD38+/H11001/H11001 expression (SSC/CD38) since CD138 expression was low in tonsil PCs and heterogeneous in peripheral PCs.

Figure 3. Phenotype of CD45bright myeloma cells. The phenotype of myeloma cells from 4 patients was determined in a 4-color assay. Myeloma cells were identified by coexpression of CD38 and CD138, and their phenotype (CD11a, CD19, CD27, CD28, CD56, Apo2.7) was analyzed in both CD45low/neg and CD45bright subsets. Ig indicates immunoglobulin.
the most proliferative compartment, could have some important consequences for tumor behavior. For example, we have previously observed that ICAM-2/LFA-1 interactions were involved in the (negative) control of myeloma cell growth through CD40.20 Interleukin 6 (IL-6) has been shown to be a survival and growth factor for human myeloma cells.21,22 More recently, it has been demonstrated that IL-6 preferentially stimulates CD45 bright myeloma cells to proliferate23,24 through activation of CD45-associated src kinase.24 Of note, CD45 bright myeloma cells express the growth fraction as defined in vivo by Drewinko et al1 more than 20 years ago. Thus, this “to be killed” population could be targeted through CD45- or CD11a-targeted therapies.

In conclusion, we have found that all patients with MM have a small proliferative compartment of myeloma cells characterized by a bright expression of CD45 and a specific expression of CD11a as well as a low Bcl-2 expression (sensitive to apoptosis). This CD45 bright:CD11a pos population of myeloma cells could constitute the growth fraction as defined in vivo by Drewinko et al12 more than 20 years ago. Thus, this “to be killed” population could be targeted through CD45- or CD11a-targeted therapies.

References

17. Huang D, O’Reilly L, Strasser A, Cory S. The anti-apoptosis function of Bcl-2 can be genetically separated from its inhibitory effect on cell cycle entry. EMBO J. 1997;16:4628-4638.
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