Pharmacologic inhibitors of PI3K/Akt potentiate the apoptotic action of the antileukemic drug arsenic trioxide via glutathione depletion and increased peroxide accumulation in myeloid leukemia cells

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Introduction

Arsenic trioxide (As$_2$O$_3$, ATO) is a clinically effective agent in the treatment of acute promyelocytic leukemia (APL). At physiologically tolerable concentrations (< 5 μM in plasma), As$_2$O$_3$ causes the degradation of the promyelocytic leukemia–retinoic acid receptor α (PML-RARα) fusion oncoprotein, expressed in the vast majority of APLs, forcing the cells to terminal differentiation and/or apoptosis. Moreover, albeit with lower efficacy, As$_2$O$_3$ also induces apoptosis in tumor cells lacking PML-RARα, including other types of leukemia and multiple myeloma, which opens the possibility for broader clinical application of this compound. Nevertheless, to be effective, this possibility requires the elaboration of strategies to increase the apoptotic action of As$_2$O$_3$ and to reduce the drug dosage to physiologically tolerable concentrations.

To date, the glutathione (GSH)–based redox system is the best known determinant of As$_2$O$_3$ sensitivity. Thus, the apoptotic action of As$_2$O$_3$ inversely correlates with the endogenous levels of GSH or GSH-associated enzymes in different leukemia cell types; and treatments that experimentally deplete or enhance the GSH content exacerbate or decrease, respectively, the As$_2$O$_3$ toxicity. Due to the multiplicity of functions of glutathione, this molecule may modulate drug toxicity by different mechanisms. This includes the scavenging of reactive oxygen species (ROSs), the production of which may be stimulated by As$_2$O$_3$, and the inactivation/detoxification of As$_2$O$_3$, either by direct binding to GSH or through a reaction catalysed by glutathione S-transferase. Hence, the modulation of intracellular GSH level appears as a promissory tool to regulate the clinical efficacy of As$_2$O$_3$.

An important aspect of apoptosis regulation is the signaling by serine/threonine kinases, a broad category of kinases that includes, among others, the mitogen-activated protein kinases (MAPKs) and the protein kinase B (PKB, Akt). Among the 3 main members of the MAPK family in mammalian cells, the extracellular signal-regulated kinases (ERK1/2) are associated to mitogenesis, and as such are generally considered as antiapoptotic. In a similar manner, the phosphatidylinositol 3-kinase (PI3K)/Akt pathway is considered as a critical survival-signaling pathway. Akt-mediated phosphorylation may alter the activity of proteins such as caspase-9, some Bcl-2 family members, and nuclear factor κB (NF-κB) and other transcription factors, which trigger or restrain apoptosis; and PI3K/Akt deregulation may contribute to tumorigenesis, metastasis, and resistance to chemotherapy. For this reason, the PI3K/Akt signaling pathway represents a promissory target of therapeutic intervention. Actually, the PI3K inhibitors...
LY294002 and wortmannin were observed to exert antitumor activity in animal cell models; and phase 1 and 2 clinical trials using rapamycin and rapamycin derivatives, which target the Akt downstream kinase mammalian target of rapamycin (mTOR), are ongoing. Of note, protein kinases and intracellular GSH are not unrelated factors. Thus, Sordet et al18 reported that the protein kinase C activator 12-O-tetradecanoylphorbol-13-acetate (TPA) potentiated the \( \text{As}_2\text{O}_3 \) toxicity by decreasing GSH content in U-937 human promonocytic leukemia cells. These observations were corroborated by us, and we also demonstrated that the TPA-provoked GSH decrease was apparently mediated by ERK activation.19 In addition, other authors demonstrated that PI3K inhibitors abrogated the insulin-provoked increase in GSH levels in rat cardiac myocytes20 and hepatocytes.21 These findings led us to examine whether GSH may represent a target of PI3K/Akt in myeloid cells, and if so, whether GSH mediates the changes in viability derived of PI3K/Akt disruption. The obtained results indicate that pharmacologic inhibitors of PI3K/Akt cause intracellular GSH depletion, increase peroxide accumulation, and potentiate the apoptotic action of \( \text{As}_2\text{O}_3 \) in a GSH-dependent manner, in U-937 promonocytes and other myeloid leukemia cell lines.

Materials and methods

Chemicals

All components for cell culture were obtained from Invitrogen (Carlsbad, CA). Monochlorobimane, dichlorohydrofluorescein diacetate (H\(_2\)DCFDA), and rhodamine 123 (R123) were obtained from Molecular Probes (Eugene, OR). DAPI (4,6-diamino-2-phenylindole) was obtained from Serva (Heidelberg, Germany). The kinase inhibitors PD98059, U0126, SB203580, SP600125, LY294002, wortmannin, and 1,6-hydroxymethyl-chiro-inositol 2-(R)-2-O-methyl-3-O-octadecylcarbonate (Akti5); the caspase-3–specific substrate N-acetyl-Asp-Glu-Val-Asp-p-nitroanilide (Ac-DEVD-pNA); the caspase inhibitor benzyl6oxy-carbonyl-Val-Ala Asp-fluoromethylketone (Z-Val-Fmk); and rabbit anti–human glutathione S-transferase PI-1 polyclonal antibody (pAb) were obtained from Calbiochem (Darmstadt, Germany). Rabbit polyclonal antibodies against human Akt, phospho-Akt (Ser\(^{473}\)), p44/42 MAPK, and phospho-p44/42 MAPK (Thr\(^{202}\)/Tyr\(^{204}\)) were obtained from Cell Signaling Technology (Beverly, MA). Mouse anti–pigeon cytomochrome \( c \) monoclonal antibody (mAb) clone 7H8.2C12, mouse anti–Bax mAb clone 6A7, and rabbit anti–rat Bcl-X pAb were obtained from BD PharMingen (San Diego, CA). Mouse anti–human Bcl-2 (100) mAb and rabbit anti–human Bax (N-20) pAb were from Santa Cruz Biotechnology (Santa Cruz, CA). Mouse anti–human 70-kDa heat shock protein (HSP70) mAb (clone C29F3A-5), which specifically recognizes the stress-inducible form of HSP70 and mouse anti–human HSP27 mAb were obtained from StressGen Biotechnologies (Victoria, BC). All peroxidase- and fluorescein isothiocyanate (FITC)–conjugated immunoglobulin G (IgG) antibodies were obtained from DAKO Diagnósticos (Barcelona, Spain). All other reagents were from Sigma (Madrid, Spain).

Cells and treatments

The human leukemia cell lines U-937 (promonocytic),22 HL-60 (myelomonocytic),23 and NB4 (acute promyelocytic)24, and Bcl-2–transfected U-937 cells (U4 clone, kindly provided by Dr Jacqueline Bréard, Institut National de la Santé et de la Recherche Médicale [INSERM] 461, Chatenay Malabry, France)25 were routinely grown in RPMI 1640 supplemented with 10% (vol/vol) heat-inactivated calf serum, 0.2% sodium bicarbonate, and antibiotics in a humidified 5% CO\(_2\) atmosphere at 37°C. For experiments, 16 to 24 hours before the initiation of the treatments the cell concentration was adjusted at approximately \( 10^5 \) cells/mL. Stock solutions of TPA (20 mM), camptothecin (10 mM), PD98059 (20 mM), U0126 (2.63 mM), SB203580 (20 mM), SP600125 (20 mM), LY294002 (20 mM), wortmannin (1 mM), Akti5 (20 mM), Z-VAD-Fmk (25 mM), monochlorobimane (200 mM), Ac-DEVD-pNA (5 mM), N-acetyl-L-cysteine (NAC, 3 M), and lonicidone (100 mM) were prepared in dimethyl sulfoxide; a stock solution of cis-platinum(II)-diammine dichloride (cisplatin, 3.3 mM) was prepared in distilled water; and a stock solution of H\(_2\)DCFDA (5 mM) was prepared in ethanol. All of these solutions were stored at \(-20^\circ\)C. Stock solutions of DAPI (10 \( \mu\)g/mL), propidium iodide (PI, 1 mg/mL), and R123 (1 mg/mL) were prepared in phosphate-buffered saline (PBS); and a stock solution of As2O3 (100 mM) was prepared in distilled water. These solutions were stored at 4°C. N-s-butylhydroxamine-5, S-sulfourea (BSO), and ascorbic acid (AA) were dissolved in distilled water at 50 and 10 mM, respectively, just before application.

Flow cytometry

The analysis of samples was carried out using an EPICS XL flow cytometer (Coulter, Hialeah, FL) equipped with an air-cooled argon laser tuned to 488 nm. The specific fluorescence signals corresponding to FITC, H\(_2\)DCFDA, and R123 were collected with a 525-nm band pass filter and the signal corresponding to PI, with a 620-nm band pass filter.

Determination of apoptosis

Distinctive characteristics of apoptotic cells were the presence of chromatin condensation/fragmentation and the acquisition of sub-G\(_1\) DNA content. To examine chromatin structure, cells were fixed with ethanol, stained with DAPI, and examined by fluorescence microscopy. To measure DNA content, cells were permeabilized, stained with PI, and examined by flow cytometry. These procedures were described in detail elsewhere.26

Measurement of caspase-3 activity

Samples of \( 4 \times 10^6 \) cells were collected by centrifugation, washed twice with ice-cold PBS, resuspended in 50 \( \mu\)L ice-cold lysis buffer (1 mM dithiothreitol, 0.03% nonidet P-40 [vol/vol] in 50 mM Tris [tris(hydroxymethyl)methoxamine, pH 7.5]), kept on ice for 30 minutes, and finally centrifuged at 14,000g for 15 minutes at 4°C. Samples containing aliquots of the supernatants (corresponding to 10 \( \mu\)g total protein), 8 \( \mu\)L caspase substrate (Ac-DEVD-pNA), and PBS to complete 200 \( \mu\)L were prepared in triplicate in 96-well microtiter plates and incubated for one hour at 37°C. The absorption was measured by spectrometry at 405 nm.

Determination of active Bax

Cells were fixed with 0.35% (vol/vol) formaldehyde and permeabilized with 0.1% (wt/vol) heat-inactivated calf serum, 0.2% sodium bicarbonate, and PBS to complete 200 \( \mu\)L. The specific fluorescence signals corresponding to FITC, H\(_2\)DCFDA, and R123 were collected with a 525-nm band pass filter and the signal corresponding to PI, with a 620-nm band pass filter.

Measurement of peroxide accumulation and mitochondrial transmembrane potential

Intracellular oxidation was measured using the fluorescent probe H\(_2\)DCFDA, which is preferentially sensitive to peroxides.24 With this aim, the cells were collected by centrifugation and incubated for one hour at 37°C in red phenol–lacking RPMI medium containing 5 \( \mu\)M H\(_2\)DCFDA, centrifuged again, and resuspended in PBS. The fluorescence was then measured by flow cytometry. To measure the mitochondrial transmembrane potential (\( \Delta \Psi \text{m} \)), the cells were incubated for 20 minutes at 37°C with PBS containing 1 \( \mu\)g/mL R123. After washing with PBS, the cells were resuspended in PBS and the fluorescence was measured by flow cytometry. Under these conditions, incubation with the depolarizing agent carbonyl cyanide \( P \)-(trifluoromethoxy) phenylhydrazone (30 \( \mu\)M) greatly decreased \( \Delta \Psi \text{m} \).
Measurement of glutathione content and glutathione S-transferase α activity

The intracellular reduced glutathione (GSH) content was currently determined by fluorometry after cell loading with monochlorobimane, following the previously described procedure. In some experiments, the content of both reduced glutathione (GSH) and glutathione disulfide (GSSG) was determined by high-pressure liquid chromatography (HPLC), according to the procedure described by Fariss and Reed, using a 5-μM Spherisorb NH₄ analytical column (Water, Milford, MA).

The glutathione S-transferase (GST) α activity was measured in cell lysates using 1-chloro-2,4-dinitrobenzene and GSH as substrates, according to the description of Jing et al.

Immunoblot assays

To obtain total cellular protein extracts, cells were collected by centrifugation, washed with PBS, and lysed by 5-minute heating at 100°C followed by sonication in Laemmli buffer containing a protease inhibitor cocktail, 10 mM sodium fluoride, and 1 mM sodium orthovanadate. To obtain cytosolic extracts (aimed at determining cytochrome c release from mitochondria), cells were collected for centrifugation; resuspended in 100 μL ice-cold PBS containing 80 mM KCl, 250 mM sucrose, and 200 μg/mL digitonin; and kept on ice for 5 minutes. After centrifugation (10,000g for 15 minutes at 4°C) the pellet was discarded. Fractions of the total or cytosolic extracts, containing equal protein amounts, were analyzed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis, blotted onto membranes, and immunodetected, as previously described.

Results

Apoptosis induction by As2O3 and other antitumor drugs, and its modulation by PI3K/Akt inhibitors

First we analyzed the capacity of As2O3, alone or in combination with the PI3K inhibitors LY294002 and wortmannin, to induce apoptosis in U-937 cells. LY294002 and wortmannin, which are known to inhibit PI3K activity with different action mechanisms, were used at 30 and 0.5 μM, respectively. These concentrations were adopted on the grounds of earlier publications, and their efficacy was here corroborated by measuring the level of phosphorylated Akt (Figure 1A, inset). It was observed that As2O3 caused a concentration-dependent (Figure 1A) and time-dependent (Figure 1B) increase in the frequency of cells with fragmented chromatin, which is characteristic of apoptosis. Some accumulation of cells at the G2 phase of the growth cycle was also detected at treatments longer than 24 hours (result not shown). Treatments for 24 hours with the PI3K inhibitors alone were innocuous (Figure 1A). Longer treatments (48-72 hours) did not cause cell death, but decreased cell proliferation by accumulating cells at the G1 phase of the growth cycle (result not shown). When used in combination, the PI3K inhibitors greatly potentiated the generation of apoptosis by As2O3, with maximum efficacy in the case of LY294002 (Figure 1A-B). These conclusions were confirmed by measuring the frequency of cells with sub-G1 DNA content (Figure 1C, and results not shown), which is also an indicator of apoptosis.

Moreover, the pan-caspase inhibitor Z-VAD-Fmk abrogated the toxicity of As2O3 alone or with LY294002 (Figure 1D), corroborating that the measured cell death is a bona fide caspase-dependent, typical apoptosis. The potentiation of As2O3-provoked apoptosis was also observed using 30 μM Akti5, a novel inhibitor that is known to directly prevent Akt phosphorylation.

To determine whether the potentiation of As2O3 toxicity by PI3K/Akt inhibitors was a cell line–specific response, experiments were carried out using human leukemia HL-60 (myelomonocytic) and NB4 (acute promyelocytic) cells. It was observed that LY294002 also potentiated the apoptotic action of As2O3 in both cell lines (Figure 1E). NB4 cells exhibited a higher sensitivity to As2O3 than U-937 and HL-60 cells, as earlier reported.

To determine whether the potentiation of As2O3 toxicity was a drug-specific response, experiments were carried out using lonidamine—an agent that, as As2O3, directly targets the mitochondria—and the DNA-specific antitumor drugs camptothecin and...
cisplatin. For homogeneity, we adopted a similar experimental design as in the case of As$_2$O$_3$, namely prolonged treatments (24 hours) with relatively low drug concentrations. It was found that, by contrast to As$_2$O$_3$, the PI3K inhibitors did not modify the apoptotic action of lonidamine, and they even decreased the apoptotic action of camptothecin and cisplatin, as measured by chromatin fragmentation (Figure 1F). This conclusion was corroborated by measuring the frequency of cells with decreased (sub-G$_1$) DNA content by flow cytometry assays, which also indicated that camptothecin and cisplatin provoked G$_2$ arrest, while lonidamine did not cause significant phase-specific blockade (results not shown).

**Protein kinase activation**

It was reported that cytotoxic agents may affect Akt phosphorylation/activation in U-937 cells. For this reason, we wanted to measure Akt phosphorylation upon treatment with As$_2$O$_3$, in the absence and presence of LY294002. Treatment with As$_2$O$_3$ alone caused a late decrease (24 hours) in Akt phosphorylation (Figure 2A). On the other hand, treatment with As$_2$O$_3$ plus LY294002 caused an earlier decrease in phosphorylation (8 hours and thereafter), with higher efficacy than treatment with LY294002 alone (Figure 2B).

Earlier publications indicated that PI3K/Akt may negatively regulate MAPK pathways. Hence, we queried whether the potentiation of As$_2$O$_3$ toxicity by PI3K/Akt inhibitors could be mediated by MAPK activation. To analyze this possibility, experiments were carried out using appropriate MAPK inhibitors, namely 10 μM PD98059 and 2.5 μM U0126 (specific for mitogen-induced extracellular kinase [MEK]/ERK), 10 μM SB20358 (specific for p38), and 10 μM SP600125 (specific for Jun N-terminal kinase [JNK]). These concentrations proved to efficaciously block kinase activation in a preceding work. It was found that SB20358 and SP20358, which were nontoxic when used alone or in combination with LY294002 (results not shown), did not affect or only minimally modified the generation of apoptosis by As$_2$O$_3$ plus LY294002 (Figure 2C), indicating that the potentiation of As$_2$O$_3$ toxicity may not be explained by a possible p38 or JNK activation. Experiments with MEK/ERK inhibitors could not be carried out since PD98059 and U0126, although nontoxic when used alone, were extremely toxic in combination with LY294002, even in the absence of As$_2$O$_3$ (result not shown). Nonetheless, immunoblot assays revealed that ERK phosphorylation was not affected by LY294002, either alone or in combination with As$_2$O$_3$, (Figure 2D), a result that apparently excludes a possible regulation via the MEK/ERK pathway.

**Modulation of mitochondria-related events**

It is known that As$_2$O$_3$ directly targets the mitochondria by interacting with the permeability transition pore. This led us to examine 2 critical mitochondria-associated regulatory events, namely the dissipation of mitochondrial transmembrane potential (∆Ψm) and the release of cytochrome c, which initiates apoptosis execution along the “intrinsic” pathway. Figure 3A shows the alteration in ∆Ψm, as measured by flow cytometry using the cationic dye R123. Treatment for 14 hours with As$_2$O$_3$ alone caused a slight ∆Ψm decrease, which was potentiated by cotreatment with LY294002. Of note, treatment with the inhibitor alone, although nontoxic (Figure 1), sufficed to slightly reduce ∆Ψm. Qualitatively similar results were obtained using the cationic dye 5,5′,6,6′-tetrachloro-1,1′,3,3′-tetraethylbenzimidazolocarbocyanine iodide (JC-1) instead of R123 (results not shown). Figure 3B shows the release of cytochrome c from mitochondria to the cytosol, as revealed by immunoblot using cytosolic extracts. As$_2$O$_3$ slightly induced cytochrome c release, which was greatly potentiated by cotreatment with LY294002. By contrast to ∆Ψm dissipation, no cytochrome c release could be detected in cells treated with LY294002 alone. The changes in cytochrome c location were followed by the activation of the executioner caspase-3, as determined by measuring DEVDase activity in cellular extracts. Thus, extracts from cells treated with As$_2$O$_3$ alone exhibited a moderate increase in DEVDAse activity, which was higher in cells treated with As$_2$O$_3$ plus LY294002 (Figure 3C). In contrast, the PI3K inhibitor did not potentiate, and instead decreased, DEVDAse activity in combination with camptothecin or cisplatin, which agrees with the changes in apoptosis (indicated in Figure 1).

The release of cytochrome c is regulated by proteins of the Bcl-2 family, which may either inhibit (eg, the antiapoptotic...
proteins Bcl-2 and Bcl-X\textsubscript{L} or promote (eg, the proapoptotic proteins Bax and Bid) the process\textsuperscript{41}. Bax remains inactive in the cytosol, and to be functional requires changes in conformation and translocation to the mitochondrial membrane\textsuperscript{27}. Bid is activated by cleavage followed by translocation to the mitochondrial membrane\textsuperscript{42}. In addition, the “heat-shock” proteins HSP27 and HSP70 also inhibit apoptosis by blocking cytochrome c and/or other elements of the intrinsic pathway\textsuperscript{43}. Hence, experiments were carried out to measure the expression and modification of these proteins. The results, represented in Figure 4, were as follows: (1) Treatment with As\textsubscript{2}O\textsubscript{3}, with or without LY294002, did not significantly modify the total Bcl-2 and Bax levels, but caused a slight decrease in Bcl-X\textsubscript{L} (Figure 4A). (2) As\textsubscript{2}O\textsubscript{3} caused Bax activation, and this activation was further enhanced by cotreatment with LY294002, as determined by flow cytometry using the 6A7 antibody. By contrast, the PI3K inhibitor did not potentiate the camptothecin-provoked Bax activation (Figure 4B). (3) LY294002 also potentiated the As\textsubscript{2}O\textsubscript{3}-provoked Bid cleavage, using as criterion the loss of the 21-kDa proform\textsuperscript{26} (Figure 4A). (4) As\textsubscript{2}O\textsubscript{3} increased HSP70 and HSP27 expression. However, while HSP70 was not affected by LY294002, the PI3K inhibitor reduced the basal HSP27 level as well as the increase caused by As\textsubscript{2}O\textsubscript{3} (Figure 4A). Hence, Bax and Bid activation, and HSP27 decrease, may contribute to the increased cytochrome c release and apoptosis execution in As\textsubscript{2}O\textsubscript{3} plus LY294002–treated cells. The possible contribution of Bad, another proapoptotic member of the Bcl-2 family (the activity of which is regulated by Akt-mediated changes in phosphorylation),\textsuperscript{16} could not be determined since phosphorylated Bad was below detection level in our immunoblot assays.

Finally, it was observed that the frequency of apoptosis was greatly reduced in Bcl-2–transfected U-937 cells (which, according to our control observations, possess an 8-fold increase in Bcl-2 content in relation to the nontransfected cells) (Figure 4C). This indicates that the generation of apoptosis by As\textsubscript{2}O\textsubscript{3} and its potentiation by LY294002 is Bcl-2 regulated, as expected in the intrinsic pathway.

**Changes in glutathione content**

As mentioned in the “Introduction,” the toxicity of As\textsubscript{2}O\textsubscript{3} is greatly dependent on the intracellular GSH content, which in turn may be affected by changes in protein kinase activities. For these reasons, determinations using the GSH-sensitive fluorescent probe monochlorobimane were carried out to analyze the possible effects of As\textsubscript{2}O\textsubscript{3} and PI3K/Akt inhibitors on intracellular GSH in U-937 cells. As indicated in Figure 5A-B, the GSH content was not reduced by treatment with 1 to 4 \(\mu\)M As\textsubscript{2}O\textsubscript{3} alone, but was considerably decreased by treatment with LY294002, either alone or in combination with As\textsubscript{2}O\textsubscript{3}. This response was corroborated using combinations of As\textsubscript{2}O\textsubscript{3} plus wortmannin or Akt\textsuperscript{5}, and using NB4 and HL-60 cells instead of U-937 cells (Figure 5B, and results not shown). The importance of GSH depletion as mediator of apoptosis...
induction in our experimental conditions was confirmed using BSO, a specific inhibitor of \( \gamma \)-glutamylcysteine synthetase (\( \gamma \)-GCS) activity, the rate-limiting enzyme of GSH biosynthesis. Preliminary determinations indicated that a 24-hour treatment with 1 mM BSO caused approximately a 60% inhibition in the intracellular GSH content (result not shown). It was observed that treatment with BSO alone was nontoxic, but as expected cotreatment with BSO increased the apoptotic action of As\(_2\)O\(_3\), in the same manner as the PI3K/Akt inhibitors. By contrast, BSO failed to increase the apoptotic action of cisplatin, camptothecin, and lonidamine (Figure 5C). This result indicates that under the conditions used here, cisplatin, camptothecin, and lonidamine behave as GSH-insensitive drugs, and may therefore explain the inability of the PI3K inhibitors to potentiate their toxicity (Figure 1F).

To shed some light on the mechanisms by which PI3K/Akt inhibition could deplete GSH, determinations were carried out using NAC, an agent earlier used as a cysteine donor for GSH biosynthesis in experiments with As\(_2\)O\(_3\).\(^{4,7}\) In addition to LY294002 and BSO, in these assays we used TPA and AA, which were also reported to reduce GSH in myeloid cells and other cell systems.\(^{4,7,18,19}\) The results are indicated in Figure 5D. NAC was unable to restore the GSH content in cells treated with BSO (which, as indicated above, blocks enzyme activity); and it was similarly ineffective in cells treated with LY294002, either alone or with As\(_2\)O\(_3\). However, NAC restored the GSH content in TPA plus As\(_2\)O\(_3\)– and AA plus As\(_2\)O\(_3\)–treated cells. This suggests that, in contrast to TPA and AA, and in the same manner as BSO, PI3K/Akt inhibitors modulate GSH biosynthesis at levels other than substrate availability. Moreover, in good parallelism with these results, NAC abrogated the increase in toxicity in the case of TPA and AA, but not in the case of LY294002 (Figure 5E), confirming again the importance of GSH as a mediator of the increased apoptosis in cells treated with As\(_2\)O\(_3\) plus PI3K inhibitors.

Final information was obtained by means of HPLC. As indicated in Table 1, this technique allowed us to corroborate the capacity of LY294002 to cause GSH depletion, and also proved that the GSH decrease provoked by the PI3K inhibitor was not adequately compensated by an increase in GSSG. Nonetheless, the ratio of GSH to GSSG was diminished, especially in As\(_2\)O\(_3\) plus LY294002–treated cells, indicating that the treatments caused GSH oxidation to some degree.

**ROS accumulation and GST\(\pi\) expression and activity**

One of the roles of GSH is the scavenging of ROSs.\(^{8}\) Hence, we queried whether the GSH depletion derived from PI3K/Akt inhibition could result in ROS overaccumulation, which might in turn explain the increase in As\(_2\)O\(_3\) toxicity. To answer this question, flow cytometry determinations were carried out using the peroxidesensitive fluorescent probe H\(_2\)DCFDA. The results are represented in Figure 6A: (1) treatment with As\(_2\)O\(_3\) alone, which as indicated above, blocks enzyme activity); and it was similarly ineffective in cells treated with LY294002, either alone or with As\(_2\)O\(_3\). However, NAC restored the GSH content in TPA plus As\(_2\)O\(_3\)– and AA plus As\(_2\)O\(_3\)–treated cells. This suggests that, in contrast to TPA and AA, and in the same manner as BSO, PI3K/Akt inhibitors modulate GSH biosynthesis at levels other than substrate availability. Moreover, in good parallelism with these results, NAC abrogated the increase in toxicity in the case of TPA and AA, but not in the case of LY294002 (Figure 5E), confirming again the importance of GSH as a mediator of the increased apoptosis in cells treated with As\(_2\)O\(_3\) plus PI3K inhibitors.

Figure 5 did not decrease GSH, did not affect peroxide accumulation; (2) treatment with LY294002 alone, which as indicated in Figures 5 and 1 sufficed to decrease GSH but was not toxic per se, increased the intracellular peroxide accumulation; and (3) a similar increase was obtained by treatment with As\(_2\)O\(_3\) plus LY294002, which as indicated in Figures 5 and 1 decreased GSH and was highly toxic.

The preceding observations could indicate that while the cells tolerate a limited ROS overaccumulation without concomitant toxicity, such overaccumulation may facilitate the generation of apoptosis by As\(_2\)O\(_3\). To investigate this possibility, experiments were carried out in which cells were cotreated with As\(_2\)O\(_3\) plus low H\(_2\)O\(_2\) concentrations (20-40 \(\mu\)M), which were per se innocuous or slightly toxic. In agreement with our hypothesis, H\(_2\)O\(_2\) potentiated or exerted a synergic effect with As\(_2\)O\(_3\), at the concentrations of 2 to 4 \(\mu\)M. By contrast, H\(_2\)O\(_2\) did not modify or exhibited only additive effects in combination with camptothecin and cisplatin (Figure 6B).

It has been indicated that GST\(\pi\) is key element in the As\(_2\)O\(_3\) detoxification machinery.\(^{11,12}\) For this reason, we asked whether the potentiation of As\(_2\)O\(_3\) toxicity by PI3K/Akt inhibitors involved an alteration in GST\(\pi\) expression and/or activity. The results in Figure 7 indicate that both the enzyme content (as measured by immuno blot assays) and activity remained unaltered or were slightly increased upon treatment with As\(_2\)O\(_3\) and LY294002, alone and in combination, when compared with the values in untreated cells.

**Table 1. Effect of treatment with LY-294002 alone (LY) and LY294002 plus As\(_2\)O\(_3\) (LY/As) on the intracellular GSH and GSSG content in U-937 cells**

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<th>Control</th>
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<th>LY/As</th>
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<td>6</td>
<td>3</td>
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<td>GSH, nmol/mg protein</td>
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<td>GSH/GSSG, nmol/mg protein</td>
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The values (nmol/mg protein) were obtained at 24 hours of treatment. For other conditions, see Figure 1 legend.

**Discussion**

The results in this work indicate that the generation of apoptosis by low concentrations of As\(_2\)O\(_3\) is potentiated by cotreatment with PI3K/Akt inhibitors (LY294002, wortmannin, Akt\(_i\),5) in U-937 promonocytic and other myeloid leukemia cells. In spite of the importance of PI3K/Akt as a survival-promoting pathway, treatment with the inhibitors alone did not significantly cause cell death, in contrast with the results sometimes obtained in other cell models.\(^{44,45}\) A possible explanation is that the intact MEK/ERK pathway provides enough survival signals to keep the cell viability.

In fact, the simultaneous inactivation of PI3K/Akt and MEK/ERKs by cotreatment with LY294002 plus either PD98059 or U0126
metabolism in myeloid cells, since the basal GSH content was reduced upon treatment with PI3K/Akt inhibitors, either alone or in combination with As$_{2}$O$_{3}$. Although our mechanistic studies on GSH regulation by PI3K/Akt are still preliminary, it may be concluded that (1) GSH depletion was not a mere consequence of cell death, since a 24-hour treatment with LY294002 alone, which was nontoxic, sufficed to considerably decrease GSH; (2) the GSH depletion was not adequately compensated by the appearance of the oxidized form, GSSG; and (3) GSH was not regulated at the level of substrate availability since (in contrast to TPA and AA, and in the same manner as BSO) the LY294002-provoked GSH depletion was not prevented by NAC. Therefore, a possible explanation is that PI3K/Akt may directly regulate γ-GCS, either at the level of enzyme synthesis or activity. Actually, PI3K inhibitors were recently reported to abrogate the insulin-mediated increase in γ-GCS catalytic subunit synthesis in rat hepatocytes, and γ-GCS activity is susceptible to modulation by changes in phosphorylation. Whatever the mechanism, GSH depletion appears to be an important factor in explaining the selective potentiation by PI3K/Akt inhibitors of apoptosis induction by As$_{2}$O$_{3}$, but not by other antitumor drugs. In fact, in contrast to As$_{2}$O$_{3}$, under the hereby assayed conditions, camptothecin, cisplatin, and lonidamine behaved as GSH-insensitive drugs, as indicated by the inability of the GSH-specific agent BSO to increase their toxicity. Due to the multiplicity of functions of glutathione, it is conceivable that different mechanisms may participate in potentiating As$_{2}$O$_{3}$ toxicity under GSH-depleting conditions. Thus, in our experiments the LY294002-provoked GSH decrease was paralleled by an increase in intracellular peroxides. This is important since ROS elevation has been reported to sensitize leukemic cells to As$_{2}$O$_{3}$-provoked apoptosis—a response here corroborated by us by cotreatment with low H$_{2}$O$_{2}$ concentrations. In addition, since GSH directly reacts with As$_{2}$O$_{3}$, GSH depletion may result in an increase in free, acting intracellular As$_{2}$O$_{3}$ concentration and hence in toxicity, even in the absence of significant alterations in GST$\gamma$ expression and activity. Of course, the GSH-based regulation does not exclude the concurrence of additional mechanisms (eg, alterations of critical cell-cycle regulatory events), which as indicated earlier in this section might explain the unusual attenuation by PI3K inhibitors of camptothecin and cisplatin toxicity, observed in this and other reports.

In summary, the present study provides the first demonstration that GSH is a target of the PI3K/Akt in myeloid leukemia cells, and as such it may partially explain the capacity of PI3K/Akt inhibitors to selectively potentiate the apoptotic action of the antileukemic drug As$_{2}$O$_{3}$. We believe that this property might provide a rationale to improve the efficacy of therapies based on the use of pharmacologic inhibitors of the PI3K/Akt pathway.

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Pharmacologic inhibitors of PI3K/Akt potentiate the apoptotic action of the antileukemic drug arsenic trioxide via glutathione depletion and increased peroxide accumulation in myeloid leukemia cells

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