Dendritic cells for NK/LAK activation: rationale for multicellular immunotherapy in neuroblastoma patients

Dominique Valteau-Couanet, Christophe Leboulaire, Kim Maincent, Muriel Tournier, Olivier Hartmann, Jean Bénard, Françoise Beaujean, Catherine Boccaccio, Laurence Zitvogel, and Eric Angevin

Natural killer (NK)/lymphokine-activated killer (LAK) cell-based immunotherapy could be beneficial against major histocompatibility complex class I-negative tumor residual disease such as neuroblastoma (NB), provided that interleukin 2 (IL-2) or surrogate nontoxic NK cell stimulatory factors could sustain NK cell activation and survival in vivo. Here we show that human monocyte–derived dendritic cells (MD-DCs) promote potent NK/LAK effector functions and long-term survival, circumventing the need for IL-2. This study demonstrates (1) the feasibility of differentiating granulocyte colony-stimulating factor–mobilized hematopoietic peripheral blood stem cells (PBSCs) into high numbers of functional MD-DCs and NK/LAK cells in a series of 12 children with stage 4 neuroblastoma (NB); (2) potent DC-mediated NK cell activation in autologous settings; (3) the reciprocal capacity of NK/LAK cells to turn immature DCs into maturing cells electively capable of triggering NK cell functions; and (4) the unique capacity of maturing DCs to sustain NK cell survival, superior to that achieved in IL-2. These data show a reciprocal interaction between DCs and NK/LAK cells, leading to the amplification of NK cell effector functions, and support the implementation of DC/NK–cell–based immunotherapy for purging the graft and/or controlling minimal residual disease after autologous stem cell transplantation. (Blood. 2002;100:2554-2561)

© 2002 by The American Society of Hematology

Introduction

Cell therapy based on autologous stem cell transplantation (ASCT) following myeloablative treatments significantly improves the prognosis of certain types of solid tumors. High-dose chemotherapy (HDC) regimen and ASCT are indicated in children with stage 4 neuroblastoma (NB).1,2 Nonetheless, tumor relapses frequently occur, either from clinically occult residual disease or from contaminating tumor cells in autologous grafts. Postgraft immunointerventions consisting of adoptive transfer of ex vivo–activated and/or antitumor effector cells derived from peripheral blood stem cells (PBSCs) in conjunction with lymphokines remain promising.

In the neuroblastoma model, several lines of evidence support the notion that antitumor immune responses might be clinically beneficial in that (1) spontaneous regressions are observed in Pepper syndrome;3 (2) tumor-infiltrating lymphocytes exhibited, in some cases, oligoclonal T-cell repertoire;4 and (3) objective responses were reported following interleukin-2 (IL-2)–systemic administration.5 However, many in vitro studies pointed to a critical susceptibility of neuroblastoma cell lines to innate effectors, that is, natural killer (NK)/lymphokine-activated killer (LAK) lysis.6,7 In the 1990s, attempts to develop clinical trials using systemic high-dose IL-2 administration with or without LAK cells after HDC plus ASCT have been hampered by severe IL-2–related toxicities reported in such children.8 Alternative strategies are clearly required to boost innate effector functions against residual neuroblastoma.

Besides their pivotal role in promoting naive T-cell activation, dendritic cells (DCs) are capable of triggering innate immune responses in vitro and in vivo. We have initially demonstrated that murine bone marrow–derived DCs (BM-DCs) promote resting NK cell activation, that is, interferon-γ (IFN-γ) secretion and dramatic cytolytic activity against tumor cells. A DC/NK cell contact is necessary and the DC-mediated triggering of NK cells is independent of IL-12 and type I IFNs.9 Interestingly, DC in vivo expansion with the use of Flt3 ligand or adoptive transfer of ex vivo–propagated DCs into palpable tumors led to NK cell–dependent antitumor effects. Whether human monocyte–derived dendritic cells (MD-DCs) could similarly promote NK cell effector functions was also recently addressed by our group and others.10-14 Human MD-DCs derived from healthy donors were shown to enhance proliferation and effector functions of resting or IL-2–activated NK cells through a bidirectional cross-talk in which activated NK cells also induce MD-DC activation. Moreover, MD-DC–mediated IFN-γ production and cytolytic activity of NK cells were dependent on cell-to-cell contacts and were independent of soluble factors, that is, IL-12, IL-15, and IL-18.

Here we analyzed the functionality and relevance of the reciprocal DC/NK cell cross-talk in a preclinical study conducted in children with NB. In a series of 12 children with stage 4 NB, we show that granulocyte colony-stimulating factor (G-CSF)–mobilized PBSCs can be differentiated into high numbers of functional MD-DCs and NK/LAK cells and that coculture of autologous DCs/LAK cells allows reciprocal activation of both subsets, long-term IL-2 deprivation, and sustained NK/LAK antitumor effector functions. These data show that in NK/LAK-based...
immunotherapy, DCs can substitute for NK cell stimulatory soluble factors, which were not devoid of toxicity.

**Patients and methods**

**Patients and sample collection**

Twelve children (2 to 7 years old) with stage 4 NB, treated according to the Société Française d’Oncologie Pédiatrique NB97 protocol were included in this study. Before initiation of treatment, all parents signed an informed consent reviewed by the Hôpital Kremlin Bicêtre ethics committee. PBSCs, collected after at least 4 courses of conventional chemotherapy to perform the ASCT, were mobilized either by chemotherapy and G-CSF (5 µg/kg/d) (Neupogen; Amgen, Neuilly sur Seine, France) or in steady state with G-CSF alone (10 µg/kg/d). Cryopreservation and thawing of cell suspensions were performed in clinical grade conditions as previously described.20 In addition, peripheral blood mononuclear cells (PBMCs) from paired blood samples were collected before and during mobilization, isolated on Ficoll density gradient, and kept frozen in liquid nitrogen until use. PBSCs contained 49.6% ± 25.3% CD14+/CD15− cells; 18.7% ± 12.6% CD3+ T lymphocytes; and 6.1% ± 3.0% CD56+CD3− NK lymphocytes. No significant difference in the repartition of these cell subsets was observed between PBSCs and the paired PBMC samples.

**Culture procedures**

Culture procedures are outlined in Figure 1. Monocyte-derived dendritic cell (MD-DC) derivation. Total leukocytes were incubated at 5 × 10^6/mL in 75-cm² culture flasks in AIM-V medium (Gibco-BRL, Paisley, United Kingdom) supplemented with 10% fetal calf serum (FCS; Pan Biotech, Aidenbach, Germany) for 2 hours at 37°C. Nonadherent cells were removed; adherent cells were washed 3 times in 1 × phosphate-buffered saline (PBS; Gibco, Paisley, United Kingdom) and cultured until day 7 in AIM-V with 10% FCS supplemented with recombinant human IL-4 (rhIL-4, 1000 U/mL; Schering Plough, Kenilworth, NJ) and recombinant human granulocyte-macrophage colony-stimulating factor (rhGM-CSF, 1000 U/mL; Leucomax, Schering Plough) in order to derive MD-DCs at an immature stage (iDCs). In some experiments, MD-DCs were induced to maturation (mDCs) with the use of lipopolysaccharide (LPS, 20 µg/mL; Sigma Chemical, St Louis, MO) added in cultures on day 5 for 48 hours.

**IL-2 activation for NK/LAK generation.** Mononuclear cells were plated in 24-well culture plates at 1 × 10^6 CD3+ and CD56+ lymphocytes per milliliter in RPMI 1640 (Gibco-BRL) supplemented with 10% human AB serum (Institut Jacques Boy, Reims, France). The rhIL-2 (1000 IU/mL, Proleukin; Chiron, Ratingen, Germany) was added only once, at day 0 of the culture.

**Coculture of MD-DCs with NK/LAK.** At day 7 or 8, IL-2-activated bulk NK/LAK cells (or immunopurified CD3+ and CD3− lymphocyte subsets) and day-7 immature autologous MD-DCs were collected, washed twice, and resuspended in supplemented RPMI 1640 medium with 10% human AB serum at a concentration of 1 × 10^6 cells per milliliter. Each cell suspension (in 0.1 mL) was plated in 96-well round-bottomed culture plates at a 1:1 iDC/NIK cell ratio. Control cultures of bulk NK/LAK cells or iDCs were also used. Immunopurifications of CD3+ and CD3− lymphocyte subsets from bulk NK/LAK cell cultures were carried out by means of magnetic cell separation after a 20-minute incubation step with an anti-CD3 monoclonal antibody (mAb) used at saturating conditions and a second incubation step with Dynabeads coated with sheep antimouse immunoglobulin G (IgG) antibodies (Dynal, Oslo, Norway) or by means of the NK cell isolation kit (Miltenyi Biotec, Bergisch-Gladbach, Germany). To dissect the DC/NK interactions, further cocultures of day-7 iDCs with or without LPS (10 µg/mL) or mDCs (as described above with the use of LPS added at day 5 for 48 hours) with 1 × 10^6 immunopurified day-7 NK/LAK cells per milliliter were performed at 3 NK/MD-DC ratios (1:1, 5:1, and 10:1). Control cultures included NK/LAK cells alone with or without IL-2 (1000 IU/mL). At days 3, 5, and 7 of coculture, viable lymphocytes were counted after trypan blue exclusion of dead cells, and the percentages of CD56+CD3− NK/LAK cells were determined by flow cytometry. Survival values were determined as the percentages of live CD56+CD3− total numbers at days 3, 5, and 7 compared with day 0 of coculture. In addition, NK/LAK cells were assayed for cytotoxicity, and IFN-γ and IL-12−p70 heterodimer production was assessed on supernatants of cocultures with the use of enzyme-linked immunosorbent assay (ELISA) kits (Immunotech, Marseilles, France).

**Immunophenotyping**

Three- and 4-color immunostainings were performed with the use of fluorescein isothiocyanate (FITC−), phycoerythrin (PE−), peridinin chlorophyll A protein (PerCP−) and allophycocyanin-conjugated mAbs (all purchased from Becton Dickinson, Pont de Claix, France) in the following combinations: day 0, PBSCs using CD14/CD56/CD3/CD45 and, in some experiments, CD15/CD14/CD5/CD56; day 7, MD-DC cultures using CD40/CD83/HLA-DR, CD86/CD1a/human leukocyte antigen-DR (HLA-DR), and CD80/CD14/HLA-DR; day 7, NK/LAK cultures using CD4/CD8/CD3/CD45, CD25/CD56/CD3/CD45, and HLA-DR/CD69/CD3/CD45; and days 3 and 7, NK/LAK with or without MD-DC cocultures using CD86/CD83/HLA-DR/CD45, CD16/CD56/CD3/CD45, and CD16/CD3/CD8/CD56. Direct stainings were performed by adding the combinations on 0.3 × 10^6 washed cells for 15 minutes at 4°C. Analyses were performed after washing twice and fixing in 1 × PBS with 0.1% paraformaldehyde. In some experiments, immunostainings of cocultures with a panel of unconjugated anti-NK receptor mAbs (all kindly given by Dr Eric Vivier, Centre d’Immunologie de Marseille-Luminy, Marseilles, France) were performed, including EB6 (anti-p58.1), GL183 (anti-p58.2), FSTR (anti-killer cell inhibitory receptor–2DS4 [anti-KIR2DS4] or anti-p58.3), DEC66 (anti-KIR3DL2 or anti-p140), CD94, ZIN270 (anti-NKG2A), 191B8 (anti-NKRP1), and BAB281 (anti-NKp46). Indirect immunostainings were performed on 0.2 × 10^6 cells for 30 minutes at 4°C. Cells were then washed twice in 1 × PBS with 0.5% bovine serum albumin and incubated with a PE-conjugated goat anti-mouse anti-IgG for 20 minutes at 4°C. After a 30-minute saturation step at 4°C in 1 × PBS with 10% mouse serum, a direct staining with CD16-FITC, CD56-cyamine-5, and CD3-alcphocyanin was performed as described above. Controls included isotype-matched immunoglobulins. Fluorescence-activated cell sorter (FACS) analysis was performed on a FACSCalibur cytometer (Becton Dickinson, BDIS, San Jose, CA) by means of Cellquest software (Becton Dickinson). Ten thousand cells were acquired according to standard forward scatter (FSC)/side scatter (SSC) criteria. Phenotypic analyses were determined within CD45+ gating for optimal exclusion of residual red cells and debris.

**Functional assays**

**Cytotoxicity assays.** The cytolytic activity of NK/LAK cell cultures was measured in a standard 4-hour 51Cr-release assay against K562 and Daudi cell line targets. In addition, the lytic activity was also tested against 2 established NB cell lines, IGR-NB1 (NB-91) and IGR-NB2 (GAU). Furthermore, the susceptibility of freshly isolated NB cells to autologous NK/LAK killing was assessed with the use of short-term NB cell lines (fewer than 3 passages and referred as AUTO NB) established from invaded bone marrow samples. Briefly, target cells (10^6) were labeled with 150 µCi

---

**Figure 1.** Outline of the experimental protocol in 12 children with a stage 4 NB. PBSCs, collected by cytopheresis, were mobilized by G-CSF or with chemotherapy and cryopreserved for ASCT. After thawing, samples were split into parallel cultures for MD-DC differentiation and NK/LAK generation, during 1 week. Cocultures of NK/LAK + MD-DCs were performed in most conditions at a 1:1 ratio for the next 7 days. Cultures were followed by immunophenotyping, cytotoxicity assay, and mixed lymphocyte reaction.
(5.55 MBq) Na$^{31}$CrO$_4$ (1 mCi/mL; [37 MBq/mL]; Dupont, Boston, MA) and $2 \times 10^5$ target cells per well were incubated with effector cells at different effector-target (E/T) ratios for 4 hours at 37°C. Experimental, spontaneous, and maximal releases of $2 \times 10^5$ labeled target cells were counted on a microplate scintillation counter (TopCount-NXT, United Technologies, Packard Laguna Hills, CA). For each E/T ratio, the percentage of lysis was calculated conventionally as follows: % lysis = (experimental cpm − spontaneous lysis cpm)/(maximal lysis cpm − spontaneous lysis cpm). All determinations were made in triplicate cultures, and data were calculated as mean ± SEM.

**Mixed lymphocyte reactions.** T-cell allostimulatory functions of immature or LPS-treated MD-DCs were assessed in triplicate cocultures of $2 \times 10^5$ allogeneic mononuclear cells with a decreased number of MD-DCs. Proliferative capacity was determined after overnight pulsing with $[^{3}H]$thymidine (1 µCi [0.037 MBq] per well; NE-NEN, Paris, France) at day 5 of the coculture. Cells were harvested onto 96-well Unifilter microplates and dried overnight, and the radioactivity was counted on a microplate scintillation counter (Topcount-NXT). All determinations were made in triplicate wells, and data were calculated as mean ± SEM.

**Results**

PBSCs from 12 children with a stage 4 NB were collected after mobilization by G-CSF with or without chemotherapy and thawed for the study with the clinical procedures used for ASCT. After the gating of mononuclear cells to exclude contaminating granulocytes, CD45$^+$ PBSCs contained 49.6% ± 25.3% CD14$^+$ CD15$^+$ cells, 18.7% ± 12.6% CD3$^+$ T lymphocytes, and 6.1% ± 3.0% CD3$^+$ CD56$^+$ NK lymphocytes. No significant difference in the repartition of these cell subsets was observed between PBSCs and the paired PBMC samples; even so, CD14$^+$ cells were systematically found enriched in PBSCs (data not shown).

**PBSCs from children with NB are a source of functional MD-DCs**

MD-DCs were derived from the adherent fractions of PBSCs by culture in AIM-V medium with 10% FCS and IL-4 plus GM-CSF for 1 week. It is noteworthy that starting CD14$^+$ cells in PBSCs were distributed in 2 phenotypes according to CD15 expression: CD14$^+$CD15$^+$ (21.6% ± 6.8% true monocytes) and CD14$^+$CD15$^+$ (78.4% ± 6.8%). From day 5, cultures contained a majority of floating veiled cells with a typical morphology of MD-DCs. At day 7, the mean MD-DC yield (mean recovery of 42.6 ± 35.1 × 10$^6$ MD-DCs) was 10.1% ± 2.1% of the starting PBSCs (367.0 ± 284.0 × 10$^6$ cells).

Differentiation, assessed by immunophenotyping at day 7, revealed that MD-DCs exhibit an immature phenotype (iDCs; CD14$^-$, HLA-DR$^{dim}$, CD40$^-$, CD80$^{dim}$, CD86$^{dim}$, CD1abright, CD83$^-$) (Figure 2A). Induction of maturation using a 48-hour LPS stimulation resulted in increased expression of HLA-DR and costimulatory molecules as well as up-regulation of CD83 (mDCs). As shown in Figure 2B, day-7 MD-DCs induced a significant proliferation of allogeneic PBMCs that dramatically increased after induction of maturation (optimal ratio of 1:5 versus 1:25, $P < .001$). Overall, regardless of the conditioning regimen, neither phenotypic nor functional differences were evidenced between MD-DCs derived from PBSCs or paired PBMC samples (data not shown).

**NK/LAK cells from IL-2–activated PBSCs display high cytotoxicity against NB tumor cells**

PBSCs and paired PBMCs were cultured in the presence of 1000 IU/mL rIL-2 at a concentration of $1 \times 10^5$/mL effectors CD3$^{+/-}$ CD56$^+$ lymphocytes. As shown in Figure 3A, phenotypic analysis displayed a significant and gradual expansion of CD56brightCD3$^-$ NK cells between day 0 and 7 (ie, 4- to 9.4-fold), representing 6.1% ± 3.0% to 31.8% ± 12.6% of CD45$^+$ cells, respectively ($P < .001$). The same range of expansion was observed in paired PBMC samples collected before (13.0% ± 5.2% to 65.2% ± 21.5%) and during (10.1% ± 7.3% to 49.0% ± 25.6%) mobilization. All together, starting with PBSCs or PBMCs, the percentage of all NK cells was increased by 5-fold. At day 7, the early and late activation markers, CD69 and HLA-DR, were significantly up-regulated on CD56brightCD3$^-$ NK cells in a mean percentage range of 58.6% ± 2.3% to 22.3% ± 10.8% of these cells, respectively (data not shown).

Lytic activity of cultures was tested at days 2, 4, and 7 against the K562 and Daudi cell lines, as well as against the 2 NB cell lines. As early as day 2, rIL-2–treated PBSCs displayed a strong cytotoxicity against K562 and Daudi (42.3% ± 14.7% and 34% ± 15.6%, respectively, at 25:1 E/T ratio). This NK/LAK activity progressively increased with culture duration with up to 50.2% ± 15.3% and 49.8% ± 14.2% lysis, respectively, at day 7, and remained high, even at a low E/T ratio (30.2% ± 9.8% and 35.8% ± 13.2% against K562 and Daudi, respectively, at a 1:5 E/T ratio). Interestingly, the 2 NB cell lines were also sensitive to lysis by activated effectors. Although lower at day 2 and 4, cytotoxicity against NB-1 and NB-2 was comparable to that achieved against K562 and Daudi at day 7 (Figure 3B). Interestingly, we confirmed the
susceptibility of freshly isolated NB cells to autologous NK/LAK killing. As shown in Figure 3C, day-7 immunopurified CD56+ NK/LAK cells lysed autologous freshly isolated NB cells from invaded bone marrow as efficiently as allogeneic targets (NB1 and DAUDI cell lines).

Taken together, PBSCs displayed a significant cytotoxicity increase on the 4 target cells lines between day 2 and day 7, \((P = .005\) and \(P = .007\), respectively, with the use of a paired \(t\) test), but not between day 2 and day 4. In addition, we did not observe major significant differences related to mobilization (data not shown), except against the Daudi target at day 4 (mean ± SD percentage lysis at 6.25:1 ratio; PBMCs before mobilization were 64.5% ± 17.0% versus PBMCs during mobilization (40.7% ± 17.7%) versus PBMCs (35.0% ± 12.0%) (PBMCs before mobilization versus PBMCs during mobilization, \(P = .101\); PBMCs before mobilization versus PBSCs, \(P < .001\); PBMCs during mobilization versus PBSCs; nonsignificant); this suggests a potential impact of G-CSF on NK functions, but this impact was not evidenced at day 7 or on other cell targets.

Overall, these results suggest that potent non–MHC-restricted cytolytic activity (against autologous and allogeneic NB tumor cells) is optimally displayed by PBSCs after 7 days of rIL-2 stimulation that is mediated mostly by activated CD56brightCD3+ NK/LAK cells.

### MD-DCs sustain survival and activation of NK/LAK cells

Because IL-2 was associated with toxicity in children but was required to maintain NK/LAK cell survival in vivo, we tested whether cocultures of MD-DCs with NK/LAK cells in the absence of exogenous rIL-2 would maintain and/or enhance LAK activity in autologous systems. Therefore, day-7 rIL-2–deprived NK/LAK cells were cocultured with day-7 immature MD-DCs derived from autologous PBSCs in 10 independent experiments from 7 NB patient samples. These cocultures, as well as control cultures of NK/LAK alone with or without IL-2, were evaluated for cell viability, immunophenotype, and cytolytic activity. Survival of activated CD56(bright)CD3− NK/LAK cells cocultured with MD-DCs was dramatically enhanced compared with control cultures (NK/LAK cells with or without IL-2) as assessed by trypan blue exclusion and flow cytometry analysis counting (Figure 4A). In addition to survival, NK/LAK cell cytotoxic activity was also significantly maintained until day 7 of coculture in the absence of IL-2. Very high levels of cytotoxicity were obtained with median percentages of lysis at a 6.25:1 E/T ratio of 56% (range, 52%-99%); 56% (range, 53%-72%); 12% (range, 9%-26%); 3% (range, 1%-23%) against K562, Daudi, NB1, and NB2 respectively (Figure 4B). Cytolytic activity was not detected when the cells were separated by a porous membrane (transwells), arguing in favor of a
direct cell-to-cell contact for the DC-mediated NK cell survival and activation (data not shown). The MD-DC–mediated NK cell activation could not be accounted for by the T/LAK component. Indeed, coculture of MD-DCs with CD3− and CD3+ cell subsets immunopurified from NK/LAK bulk suspensions demonstrated that the responding effectors were indeed CD56+CD3− and not CD3+ cells (Figure 4B, inset). Therefore, MD-DCs selectively activate NK/LAK cells in vitro in a cell-to-cell contact–dependent manner.

The phenotypic characterization of NK/LAK cells cocultured with MD-DCs included CD16+/CD3−/CD56−/CD45− immunostainings. Before coculture, on the basis of CD16 density, 3 CD56bright subsets were identified, CD56brightCD16−, CD56brightCD16dim, and CD56brightCD16bright cells, while most CD56dim cells expressed CD16 at high density. Interestingly, at day 7, the percentage of CD56brightCD16+ NK/LAK cells surviving in coculture with DCs was similar to that observed prior to IL-2 deprivation, while residual viable NK/LAK from control cultures deprived of IL-2 were mostly CD56bright CD16bright cells, while most CD56dim cells expressed CD16 at high density. Interestingly, at day 7, the percentage of CD56brightCD16+ NK/LAK cells surviving in coculture with DCs was similar to that observed prior to IL-2 deprivation, while residual viable NK/LAK from control cultures deprived of IL-2 were mostly CD56bright CD16bright with a dramatic clearance of mature CD56dim cells (Figure 4C). Overall, this phenotype was associated with high levels of cytotoxicity displayed by the cocultures. However, such CD56brightCD16+ cells did not up-regulate NKRP1- or Nkp46 (BABB281)–activating receptors nor did they down-regulate CD158a/p58.1, CD158b/p58.2, CD94/NKG2A, KIR2DS4/p58.3, or KIR3DL2/p140 inhibitory receptors following interaction with MD-DCs. All together, autologous immature MD-DCs maintain survival and effector functions of CD56bright/CD16+ NK/LAK cells in the absence of exogenous IL-2.

**LPS-activated MD-DCs are the most potent antigen-presenting cells (APCs) for NK/LAK activation**

To further dissect the dynamic of the DC/NK cell cross-talk, cocultures of CD56+CD3− immunopurified day-7 NK/LAK cells were performed with MD-DCs at various stages of maturation, that is, immature MD-DCs (iDCs; CD86dim/CD83−), maturing DCs (iDCs with LPS stimulation), or mature MD-DCs (mDCs; CD86bright/CD83+) after 2 days of LPS stimulation, at various NK/DC ratios (1:1, 5:1, 10:1). NK cell functions were assessed (cytotoxicity, IFN-γ production) following 3- to 7-day cocultures. Interestingly, at all differentiation stages, DCs can maintain (day 3; Figure 5A) and promote (day 7; Figure 5B) NK cell activation. This DC-mediated NK cell activation even occurred at a 10:1 NK/DC ratio and, most strikingly, for maturing DCs (Figure 7B,E). In contrast, mature DCs did not promote NK cell survival (26%, 27.8%, and 41.8% of CD45+/CD16− NK/LAK cell survival at days 3 and 7, respectively). As expected, mDCs seemed to be engaged toward whether NK/LAK cells could activate such iDCs. Indeed, a significant up-regulation of CD86 and CD83 molecules was observed in the whole CD45+ HLA-DR+ population (Figure 6A, center panels), as compared with iDCs without NK cells (Figure 2A). Up to 33% of iDCs became CD83+/CD86bright in the presence of NK/LAK cells, a percentage that was further enhanced (56%) in the maturing DCs (iDCs with LPS in the coculture). As already reported, it is likely that iDCs represented NK cell targets, since we observed 26%, 27.8%, and 41.8% of CD45+ HLA-DR+ cells representing iDCs, iDCs with LPS, and mDCs, respectively (DC1 + DC2 in left panels, Figure 5A) alive at day 3 of cocultures. Recruitment of multiple NK/LAK cells onto single iDCs could be observed at day 7 (Figure 7A).

Interestingly, the most activating DCs for NK/LAK cells, that is, maturing DCs (iDCs with LPS), were also those that significantly promoted NK cell long-term survival without exogenous IL-2 (up to 25%; Figure 6B, left panels). Morphologic analyses highlighted clustering of DCs with NK/LAK cells, mostly in the maturing DCs (Figure 7B,E). In contrast, mature DCs did not promote NK cell survival (27% and 2% survival at days 3 and 7, respectively).

**Reciprocal cross-talk between MD-DCs and activated NK/LAK cells**

Because immature DCs were also capable of maintaining long-term activation of NK/LAK cells, we addressed the question of
apoptotic process (Figure 7C). It is noteworthy that most NK cells alive at day 7 exhibited a mature phenotype, CD56^bright/CD16^bright, that is associated with effector functions.

All together, NK/LAK cells can directly activate immature DCs with or without additional inflammatory stimuli, and such activated DCs, in return, are empowered to maintain not only NK cell effector functions but also long-term survival.

Discussion

The purpose of this study was to investigate the potential therapeutic relevance of the newly described DC/NK cell cross-talk on the basis of our previous results demonstrating that mouse DCs enhance NK cell functions in vitro and in vivo, and on recent similar reports in human in vitro model systems demonstrating that DCs can act on the priming phase of NK cell activation. The purpose of this study was to investigate the potential therapeutic relevance of the newly described DC/NK cell cross-talk on the basis of our previous results demonstrating that mouse DCs enhance NK cell functions in vitro and in vivo, and on recent similar reports in human in vitro model systems demonstrating that DCs can act on the priming phase of NK cell activation.

DC/NK CROSS-TALK IN NEUROBLASTOMA

DCs can act on the priming phase of NK cell activation. The purpose of this study was to investigate the potential therapeutic relevance of the newly described DC/NK cell cross-talk on the basis of our previous results demonstrating that mouse DCs enhance NK cell functions in vitro and in vivo, and on recent similar reports in human in vitro model systems demonstrating that DCs can act on the priming phase of NK cell activation.

Discussion

The purpose of this study was to investigate the potential therapeutic relevance of the newly described DC/NK cell cross-talk on the basis of our previous results demonstrating that mouse DCs enhance NK cell functions in vitro and in vivo, and on recent similar reports in human in vitro model systems demonstrating that DCs can act on the priming phase of NK cell activation.
exhibited an immature phenotype in that they were CD83+ and strongly responded to inflammatory stimuli for up-regulation of costimulatory molecules and alloreactivity. In respect to our end point, the proof of the principle of consistent propagating functional DCs for NK/LAK activation, there was no significant difference in rates or functions with the use of steady-state versus mobilized PBSCs for DC propagation by means of the process of adherence.

IL-2 has been administered in vivo to expand and/or sustain LAK activity of NK/LAK cells propagated at high numbers ex vivo. However, the selective expansion of human NK cells during continuous in vivo infusion of low-dose IL-2 probably results from enhanced NK cell differentiation from CD34+ hematopoietic progenitors combined with an IL-2-dependent delay in NK cell death, rather than from proliferation of mature NK cells in the periphery.24 A similar observation has been recently reported with the newly identified IL-21 cytokine.25 Other NK cell–stimulatory factors, IL-12, tumor necrosis factor–α (TNF-α), IL-1β, IL-10, or cytokines known to use IL-2 receptor-γ (IL-2Rγ) common chain for signaling (IL-4, IL-7, IL-9, IL-13), were not shown to significantly sustain NK cell survival in vitro. Only IL-15, much more abundant in normal tissues than IL-2, was demonstrated to prolong NK cell survival at picomolar concentrations in vitro, but it is not available for clinical grade infusions.26 We previously demonstrated in a mouse model that BM-DCs promote resting NK cell effector functions in vitro and mediate NK cell–dependent tumor rejection in Rag2−/− mice.27 Recent studies28,29 have shown that human MD-DCs enhance proliferation and effector functions of both resting and IL-2–activated NK cells. The DC-mediated NK cell activation relies on cell-to-cell contacts with no substantial contribution of IL-12, IL-15, or IL-18 for the NK cell effector functions. Now, we implement these findings to autologous systems and demonstrate that MD-DCs are able to maintain the survival of 7-day NK/LAK cells for a week in the absence of exogenous cytokines. Importantly, cytolytic activity of NK/LAK cells in coculture with DCs remained potent against K562, Daudi, and NB cell lines. Such IL-2–deprived 7-day NK/LAK cells in coculture with DCs were mostly CD56bright/CD16− or CD16+ with loss of the mature CD56dim counterparts but not T cells.

We further dissected the dynamics of the DC/NK cell cross-talk in these autologous settings by studying the effects of DC-differentiation stage on NK cell effector functions and survival. Strikingly, immature DCs maturing under inflammatory stimulation or NK/LAK signaling were the most potent antigen-presenting cells triggering cytosis, IFN-γ production, and survival of NK/LAK cells. In contrast, mature DCs were unable to trigger IFN-γ production or sustained NK cell survival in such conditions. Importantly, cell-to-cell contact was critical for DC-mediated NK cell activation and was even more potent than rhuIL-2. These findings could be relevant in the early steps of innate immune response. Tissue-resident DCs encountering pathogens or tumor cells might undergo an activation process that will concomitantly recruit blood-resting NK cells, favoring maturing DC/NK cell interaction. The DC/NK cell cross-talk that is initiated will likely lead to NK cell triggering and have the result that activated NK cells will favor clustering and activation of newly recruited DCs at
the site of inflammation; in addition, when the NK/DC ratio favors NK cells, NK cells become NK cell targets to shut off the process.

Regulatory pathways of recruitment and migration of NK cells remain largely unknown. In mice infected with tumor cells, it has been recently shown that tumor cells lacking appropriate MHC class I expression induced NK cell inhibition, cytotoxic activation, and induction of transcription of γ-IFN in KIR-bearing NK cells. It is conceivable that reinfused NK/LAK cells could traffic to NB tumor deposits because such tumor cells are often MHC class I-negative. Interestingly, NK cells and DCs are known to be responsive to similar chemokines, such as macrophage-inhibitory protein-3x in inflamed sites and virus-infected mice, where NK cells have been found in close contact with splenic APCs. This interaction could be pivotal for a chemokine-to-cytokine-to-chemokine cascade leading to local control of the inflammation; in addition, when the NK/DC ratio favors NK cells, NK cells become NK cell targets to shut off the process.

These results support the use of PBSCs not only as a hematopoietic support but also as a source of effector cells with additive or synergistic functions. The ability to generate both NK/LAK effectors and MD-DCs in short-term cultures dramatically extends the therapeutic field of autologous PBSCs. The administration of pluriopotent grafts might improve antitumor activity in vivo and lessen IL-2–related morbidity. NB constitutes an ideal tumor model to demonstrate the efficacy of such a combined multicular adoptive therapy.

Acknowledgments

We are indebted to Dr Chantal Le Forestier from ETS Créteil for providing PBSC samples and to Gwenaëlle Leroux from Institut Gustave Roussy for expert technical assistance in recovery and culture of neuroblastoma cells. We also thank Dr Eric Vivier for helpful discussions.

References

17. Miller JS, Prosper F, McCullar V. Natural killer (NK) cells are functionally abnormal and NK cell progenitors are diminished in granulocyte colony-stimulating factor-mobilized peripheral blood progenitor cell collections. Blood. 1997;90:3098-3105.

From www.bloodjournal.org by guest on October 17, 2017. For personal use only.
Dendritic cells for NK/LAK activation: rationale for multicellular immunotherapy in neuroblastoma patients

Dominique Valteau-Couanet, Christophe Leboulaire, Kim Maincent, Muriel Tournier, Olivier Hartmann, Jean Bénard, Françoise Beaujean, Catherine Boccaccio, Laurence Zitvogel and Eric Angevin